

JÚLIA JANTSCH FERLA

**INTERCROPPED PLANTS AS A RESERVOIR OF PREDATORY MITES IN
COFFEE CROP WITH A DESCRIPTION OF A NEW SPECIES**

Dissertação apresentada à Universidade Federal de Viçosa, como parte das exigências do Programa de Pós-Graduação em Entomologia, para obtenção do título de *Magister Scientiae*.

Orientador: Angelo Pallini Filho

Coorientadores: André Lage Perez
Gustavo Júnior de Araújo
Madelaine Venzon
Noeli Juarez Ferla

**VIÇOSA - MINAS GERAIS
2021**

**Ficha catalográfica elaborada pela Biblioteca Central da Universidade
Federal de Viçosa - Campus Viçosa**

T
Ferla, Júlia Jantsch, 1995-
F357i Intercropped plants as a reservoir of predatory mites in
2021 coffee crop with a description of a new species / Júlia Jantsch
Ferla. – Viçosa, MG, 2021.
1 dissertação eletrônica (66 f.): il. (algumas color.).

Texto em inglês.
Orientador: Angelo Pallini Filho.
Dissertação (mestrado) - Universidade Federal de Viçosa.
Inclui bibliografia.
DOI: <https://doi.org/10.47328/ufvbbt.2021.144>
Modo de acesso: World Wide Web.

1. Tydeidae - Controle biológico. 2. Serviços ambientais.
3. Tydeidae - Classificação. 4. Café - Doenças e pragas -
Controle biológico. I. Universidade Federal de Viçosa.
Departamento de Entomologia. Programa de Pós-Graduação em
Entomologia. II. Título.

CDD 22. ed. 632.6542

Bibliotecário(a) responsável: Alice Regina Pinto CRB6 2523

JÚLIA JANTSCH FERLA

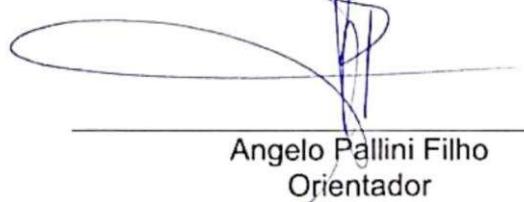
**INTERCROPPED PLANTS AS A RESERVOIR OF PREDATORY MITES IN
COFFEE CROP WITH A DESCRIPTION OF A NEW SPECIES**

Dissertação apresentada à Universidade Federal de Viçosa, como parte das exigências do Programa de Pós-Graduação em Entomologia, para obtenção do título de *Magister Scientiae*.

APROVADA: 27 de agosto de 2021.

Assentimento:


Júlia Jantsch Ferla
Autora


Angelo Pallini Filho
Orientador

AGRADECIMENTOS

Agradeço a Universidade Federal de Viçosa e ao Programa de Pós-graduação em Entomologia pela infraestrutura que permitiu que o trabalho fosse realizado e pela oportunidade de crescimento e realização profissional.

Agradeço a CAPES pela concessão da bolsa e ao Consórcio EMBRAPA/CAFÉ pelo apoio financeiro na pesquisa.

Agradeço ao Laboratório de Acarologia (LABACARI) da Universidade do Vale do Taquari – UNIVATES, onde o trabalho foi parcialmente desenvolvido.

Agradeço a Empresa de Pesquisa Agropecuária de Minas Gerais - EPAMIG e a Fazenda Experimental de Patrocínio pela infraestrutura disponibilizada para o desenvolvimento do projeto.

Agradeço ao meu orientador Angelo Pallini por todos os ensinamentos e contribuições para o meu crescimento pessoal e profissional.

Agradeço a minha coorientadora Madelaine Venzon e aos colegas do laboratório de controle biológico da EPAMIG, em especial a Elem Martins, pelo auxílio no meu trabalho e pela coleta do material.

Agradeço aos meus coorientadores André Lage Perez, Gustavo Júnior de Araújo e Noeli Juarez Ferla por todas as sugestões e contribuições.

Agradeço ao Professor Arne Janssen pelos comentários e críticas construtivas ao trabalho.

Agradeço ao Gabriel Lima Bizarro, Wesley Borges Wurlitzer, Tairis da Costa e Marcello De Giosa pela ajuda na identificação de Tenuipalpidae, Tydeidae, Tarsonemidae e Stigmeidae.

Agradeço ao Professor Enrico de Lillo por me ensinar sobre a taxonomia dos Eriophyoidea.

Agradeço aos amigos do laboratório André Costa Cardoso, Pedro Hermano Marques Gonçalves Nascimento, Milena Kalile, Shauanne Dias Pancieri, Ítalo Marcossi, Vanessa Farias da Silva e Jose Morales por toda colaboração intelectual e braçal.

Agradeço a minha turma da entomologia (os meus “Entomofriends”) que tornaram todo esse período mais leve e divertido.

Agradeço as minhas amigas do Rio Grande do Sul Jéssica Bersch, Marine Matte, Priscila Rode e Tairis da Costa por todo carinho, conversa, apoio, amizade e companheirismo.

E agradeço especialmente a minha família, que sempre me apoiou, incentivou e acreditou no meu potencial. Ao meu pai Juarez, por ter me mostrado e inserido no maravilhoso mundo da acarologia, a minha mãe Josélia por me inspirar a ser uma mulher forte como ela, e a minha irmã Laura, por estar comigo em todos os momentos e por ser minha melhor amiga.

RESUMO

FERLA, Júlia Jantsch, M.Sc., Universidade Federal de Viçosa, agosto de 2021. **Plantas consorciadas como reservatório de ácaros predadores na cultura do café, com a descrição de uma nova espécie.** Orientador: Angelo Pallini Filho. Coorientadores: André Lage Perez, Gustavo Júnior de Araújo, Madelaine Venzon e Noeli Juarez Ferla.

A taxonomia descreve e identifica táxons, de modo a permitir que uma ampla diversidade de pesquisas seja desenvolvida com base na utilização de informações taxonômicas para arquitetar estratégias de manejo de pragas. O cultivo de café no Brasil se configura como um setor de grande importância econômica para o país, além de consistir em um dos exemplos de uso da identificação de organismos associados. Atualmente o Brasil é o maior produtor de café e o sistema convencional desse cultivo é feito a partir da monocultura com baixa diversificação de plantas e do uso de insumos químicos para o controle de pragas. Os químicos utilizados afetam não somente a praga alvo, diversos artrópodes e patógenos, mas também outros organismos benéficos que desempenham importantes papéis na manutenção do ecossistema, já que estes são fornecedores de serviços ecossistêmicos, como a polinização e o controle biológico. Assim, o controle biológico conservativo pode ser usado como uma estratégia alternativa ao uso de agroquímicos quando se objetiva controlar pragas nos agroecossistemas. O uso de práticas desse controle pode ser feito com uma variedade de estratégias integradas, que utilizam os serviços ecológicos da biodiversidade local. Uma das estratégias que pode ser empregada é o plantio de espécies vegetais que fornecem abrigo e alimento alternativo, como pólen e néctar, a ácaros predadores. Esse sistema resulta no aumento da biodiversidade no agroecossistema e pode permitir que um controle estável de pragas seja alcançado. Eu estudei a comunidade de ácaros sob a perspectiva taxonômica existente em um sistema de café consorciado intercalado em uma faixa lateral com plantas que fornecem pólen e néctar para a comunidade de artrópodes. Meu objetivo foi avaliar se as comunidades de ácaros fitófagos e predadores presentes nos cafeeiros sofrem modificações conforme se aumenta a distância das Espécies de Plantas Consorciadas (EPC). Eu também investiguei se existe alteração na composição das espécies de ácaros predadores e fitófagos nas EPC e avaliei o papel que essas plantas

desempenham como estratégia de controle biológico conservativo. Além disso, eu descrevi de uma nova espécie de Tydeidae coletada em folhas de café e *Inga*. O estudo foi realizado na Fazenda Experimental da EPAMIG, em Patrocínio – MG. Os ácaros foram coletados nas EPC e em café, em diferentes distâncias em um transecto de 16 metros (4, 8, 12 e 16 metros). Para realizar a descrição da nova espécie, a identificação foi feita através de chaves dicotômicas e microscópio óptico com contraste de fases e os desenhos foram feitos com o programa Adobe Illustrator®. Nesse trabalho, eu demonstrei que as EPC avaliadas não modificam a comunidade de predadores e fitófagos ao longo de um gradiente de distância. Apesar disso, estas abrigam ácaros herbívoros, que não são pragas do café e que podem ser presas alternativas para aumentar a população de predadores e, portanto, auxiliar no controle biológico de ácaros pragas no cultivo de café.

Palavras-chave: Controle biológico conservativo. Diversificação do agroecossistema. Serviços ecossistêmicos. Tydeidae. Taxonomia.

ABSTRACT

FERLA, Júlia Jantsch, M.Sc., Universidade Federal de Viçosa, August, 2021. **Intercropped plants as a reservoir of predatory mites in coffee crop with a description of a new species.** Adviser: Angelo Pallini Filho. Co-advisers: André Lage Perez, Gustavo Júnior de Araújo, Madelaine Venzon and Noeli Juarez Ferla.

Taxonomy describes and identifies species allowing different investigation to be developed from it, becoming possible to use taxonomic information in order to build strategies for pest management. This is the case of coffee crops in Brazil, which are of great importance to the country's economy. Currently, Brazil is the world main coffee exporter and the conventional system production consists in a monoculture with low plant diversification that uses chemical inputs to control pests. This system not only affects the target pest species, different arthropods and pathogens but also many other beneficial species that can play important roles in maintaining the ecosystem due to the services they provide, such as pollination and biological control. Hence, the conservation biological control can be an alternative to pesticides while controlling insect and mite pests in agroecosystems. The use of conservation biological control practices could be done with a variety of integrated strategies using ecological services from the local biodiversity. One strategy is using plants that provide shelter and alternative food to predatory mites, such as nectar and pollen, increasing biodiversity in agroecosystems in order to reach a sustainable control of pests. I studied the mite community from a taxonomic perspective in a coffee system intercropped interspersed on a side strip with plants that provided nectar and pollen for the arthropod community. My aim was to evaluate if the communities of predators and phytophagous mites present on coffee crops changed if their distance from the selected Intercropped Plants Species (IPS) increased. I also investigated the composition of predatory and phytophagous mite species on the IPS and assessed the role of these plants as a conservative biological control strategy. Finally, I described a new Tydeidae species sampled on coffee and *Inga* plants. The study was carried out at the EPAMIG Experimental Farm, in Patrocínio county, Minas Gerais state. To do so, the mites were recorded on IPS and in different distances on coffee transects of 16 m (4, 8, 12 and 16 meters) extending from IPS. Morphological identification of the new tydeid species was

made with a microscope with phase contrast and dichotomous key and drawings were made using Adobe Illustrator® program. I show that the IPS do not modify the community of predators and phytophagous mites in coffee along the distance, but they harbor herbivorous mites that are not pests on coffee. Those mites can be an alternative prey to increase the population of predators on these plants, assisting in the biological control of mite pests in the coffee crops.

Keywords: Conservative biological control. Agroecosystem diversification. Ecosystem service. Tydeidae. Taxonomy.

SUMÁRIO

General introduction	10
References	13
Chapter 1: Intercropped plants as a reservoir of predatory mites in coffee crop	19
Abstract	19
Introduction.....	20
Material and methods.....	22
Study area.....	22
Data collection.....	23
Statistical analyses.....	24
Results	25
Abundance and richness on coffee and IPS	25
Diversity and composition on coffee crops	25
Discussion	26
Acknowledgments	29
References	29
Chapter 2: New tydeid mite (Acarí: Prostigmata) associated to coffee and inga plants.	51
Abstract	51
Introduction.....	52
Material and Methods.....	52
Results	53
Systematic.....	53
General diagnosis	53
Differential diagnosis.....	55
Etymology	55
Type material	55
Acknowledgements	57
References	58
General conclusions.....	66

GENERAL INTRODUCTION

Taxonomy describes and identifies taxons allowing different investigation to be developed from it. So, it is important to create dichotomous key or other strategies to disseminate and facilitate identification of species, which will facilitate studies on a broad range of research from basic ecology to more applied field as biological control (Moraes 1987). Once a species is identified, it is possible making taxonomic comparison of populations, identifying herbivorous pest and their natural enemies (Moraes 1987). Furthermore, when investigating arthropod communities much can be used from the taxonomic information in order to build strategies to manage pests and this is the case of coffee crops in Brazil that have great importance to the country's economy (Pallini 1991; Spongoski et al. 2005; Mineiro et al. 2006a; Marchetti 2007; da Silva et al. 2020; Conab 2021).

Currently, Brazil is the main producer and the largest exporter in the world and the species most cultivated in the country is *Coffea arabica* L. (Rubiaceae) (Conab 2021). The covered area is 1.75 million hectares, from which Minas Gerais State has the largest cultivated area (1.22 million hectares). This corresponds to 71.7% of the area occupied with this plant species nationwide, followed by Espírito Santo, São Paulo and Bahia (Conab 2021). In general, Minas Gerais is divided into three main coffee producer regions: South of the State, Zona da Mata (reminiscent of Atlantic Forest area) and Alto Paranaíba (Cerrado biome) (Conab 2021). The Cerrado biome in this state is characterized by mechanized coffee production (Conab 2020) and Zona da Mata and South of Minas Gerais has predominantly family farming production (Coelho 2005).

This crop suffers from diseases and pests that can cause economic yield loss (Le Pelley 1973; Pallini 1991; Souza et al. 1998; Spongoski et al. 2005). Some of the most important insects that cause economic losses and are serious pests in coffee agroecosystems are the coffee leaf miner *Leucoptera coffeella* (Guérin-Mèneville) (Lepidoptera: Lyonetiidae) and the coffee berry borer *Hypothenemus hampei* (Ferrari) (Coleoptera: Curculionidae: Scolytinae) (Le Pelley 1973; Souza et al. 1998). Apart from insets, there are phytophagous mites that cause economic damages such as *Oligonychus ilicis* (McGregor) (Tetranychidae) that causes defoliation, premature leaf

drop and reduction in plant photosynthesis (Reis 2005; Reis and Souza 1986) and *Polyphagotarsonemus latus* (Banks) (Tarsonemidae) that damages the leaves, curling it downward (Amaral 1951; Chagas 1973; Flechtmann 1967; Jeppson et al. 1975; Matiello 1987; Peña and Bullock 1994).

There are also some species of the genus *Brevipalpus*, that despite being of great economic importance on coffee culture, also represent economically damage to other cultures in the world (Childers and Rodrigues 2011; Nunes et al. 2018; Mineiro and Sato 2019). They can transmit viruses to many plant species and acting as vectors of the Coffee ringspot virus (Chagas et al. 2003; Childers et al. 2003; Kitajima et al. 2003; Nunes et al. 2018; Mineiro and Sato 2019). The taxonomy of this group is complex (Beard et al. 2015) and *Brevipalpus phoenicis* (Geijskes) comprises several cryptic species (Beard et al. 2013; Navia et al. 2013; Beard et al. 2015). Beard et al (2015) reorganized this group dividing one species (*B. phoenicis*) into a complex of species. Before this organization, *B. phoenicis* was recorded on coffee and after this revision there are records of *B. yothersi* and *B. papayensis* from this crop and both can transmit Coffee ringspot virus (Spongoski et al. 2005; Mineiro et al. 2006a; Mineiro et al. 2006b; Marchetti 2007; Mineiro et al. 2008; Mineiro et al. 2009; Nunes et al. 2018; Mineiro and Sato 2019). The family of predatory mites commonly found on coffee plants in Brazil are Phytoseiidae, Ascidae, Cheyletidae, among others (Pallini 1991; Spongoski et al. 2005; Marchetti 2007). Some species of this families such as Phytoseiidae are associated and feed on phytophagous mites including some of this pest families, other arthropods and also on pollen and nectar (McMurtry and Croft 1997; Croft et al. 2004; McMurtry et al 2013)

Conventional coffee system is based on monoculture with low plant diversity and the use of agrochemicals to control pests (Thrupp 1990; Cardoso et al. 2001). There are side effects in applying pesticides such as pest resistance and resurgence and the residues remaining in the field and on the fruits, that impacts biodiversity, environment and the human life (Thrupp 1990; Perfecto et al. 1996; Perfecto and Vandermeer 2008; Khan et al. 2009; Pedlowski et al. 2012; Morris et al. 2013; Sales et al. 2013; Avelino et al. 2018; Aparecido et al. 2020). Nonetheless, there are some alternatives to conventional practices in order to control pests in the crop being one of them the conservative biological control, which is the enhancement of the natural

enemies present in the agroecosystem (Letourneau et al. 2011; Ratnadass et al. 2012; Tixier 2018). To do so, a strategy is to attract and maintain the natural enemies in the system increasing diversity of plants that provide food and shelter for them (Rezende et al. 2014, 2021; Rosado et al. 2021).

Some farmers have already adopted the coffee crop intercropped system with native and exotic plant species to biome in Brazil (Souza et al. 2010). This intercrop favors to food security and production, in addition to low cost beneficial in the agroecosystem (Souza et al. 2010). In the studied system, to increase the diversity, some endemic plants from Brazil were planted intercropped with coffee as *Senna macranthera* (Dc. ex collad.) H.S. Irwin & Barnaby, *Inga edulis* Mart and *Varronia curassavica* Jacq. (Cordiaceae). *Senna macranthera* and *I. edulis* are trees with extrafloral nectaries (Koptur 1994; Melo et al. 2010) and *Varronia curassavica* is an aromatic perennial shrub that provides pollen during the whole year (Brandão et al. 2015). Together in the system, they can be a good strategy for conservative biological control by providing alternative food to natural enemies. Furthermore, there are some studies associating *S. macranthera* and *Inga* spp. with coffee crops. However, they involve mainly insects but not mites (Cardoso et al. 2001; Teodoro et al. 2009; Souza et al 2010; Rezende et al. 2014,2021).

Thus, given the importance of increasing biodiversity in agroecosystems in order to reach a sustainable control of pests, the general objective of this work was to evaluate the mite community in a coffee system intercropped with plants that provide nectar and pollen to the arthropods. To do so, in the first chapter I sampled and identified the mites in this system to evaluate if the communities of predators and phytophagous mites present on coffee crops changed if their distance from the selected intercropped plants increased. Additionally, I assessed the composition of predacious and phytophagous species on these selected intercropped plants to investigate if they are reservoir of beneficial or detrimental mites for the coffee crop.

In the second chapter, I described a new species of Tydeidae based on specimens collected on both *C. arabica* and *I. edulis*, named *Tydeus mineirensis* sp. nov. as a tribute for the first Tydeidae species described from Minas Gerais state. Additionally, given the importance of properly identification and to facilitate future

studies involving tydeids in coffee, I provided a dichotomous key of Tydeoidea reported from coffee crops in the states of Minas Gerais and São Paulo, Brazil.

References

- Amaral JF (1951) A infestação de ácaros nos cafezais. Biol 17 pp 130.
- Aparecido LEO, Rolim GS, Moraes JRSC, Costa CTS, de Souza PS (2020) Machine learning algorithms for forecasting the incidence of *Coffea arabica* pests and diseases. Int. J. Biometeorol. 64: 671–688. <https://doi.org/10.1007/s00484-019-01856-1>
- Avelino J, Allinne C, Cerda R, Willocquet L, Savary S (2018) Multiple-disease system in coffee: From crop loss assessment to sustainable management. Annu. Rev. Phytopathol. 56: 611-635. <https://doi.org/10.1146/annurev-phyto-080417-050117>
- Beard JJ, Ochoa R, Bauchan GR, Trice M, Redford A, Walters T, Mitter C (2013) Flat mites of the world. Available from: <http://idtools.org/id/mites/flatmites/> (Accessed on August 2021)
- Beard JJ, Ochoa R, Braswell WE, Bauchan GR (2015) *Brevipalpus phoenicis* (Geijskes) species complex (Acari: Tenuipalpidae) --a closer look. Zootaxa 3944: 1-67. <http://dx.doi.org/10.11646/zootaxa.3944.1.1>
- Brandão DS, Mendes ADR, Santos RR, Rocha SMG, Leite GLD, Martins ER (2015) Biologia floral e sistema reprodutivo da erva-baleeira (*Varronia curassavica* Jacq.). Rev. Bras. Pl. Med. 17: 562-569. [10.1590/1983-084X/14_011](https://doi.org/10.1590/1983-084X/14_011)
- Cardoso IM, Guijt I, Franco FS, Carvalho AF, Neto PF (2001) Continual learning for agroforestry system design: university, NGO and farmer partnership in Minas Gerais, Brazil. Agric. Syst. 69: 235-257. [https://doi.org/10.1016/S0308-521X\(01\)00028-2](https://doi.org/10.1016/S0308-521X(01)00028-2)
- Chagas CM (1973) Associação do ácaro *Brevipalpus phoenicis* (Geijskes) à mancha anular do cafeeiro. Biol 39: 229-232.
- Chagas CM, Kitajima EW, Rodrigues JCV (2003) Coffee ringspot virus vectored by *Brevipalpus phoenicis* (Acari: Tenuipalpidae) in coffee. Exp. Appl. Acarol. 30: 203–213. <https://doi.org/10.1023/B:APPA.0000006549.87310.41>

Childers CC, Rodrigues JCV (2011) An overview of *Brevipalpus* mites (Acari: Tenuipalpidae) and the plant viruses they transmit. Zoosymposia 6: 180–192. 10.11646/ZOOSYMPOSIA.6.1.28

Childers CC, Rodrigues JCV, Derrick KS et al (2003) Citrus leprosis and its status in Florida and Texas: past and present. Exp. Appl. Acarol. 30: 181-202. <https://doi.org/10.1023/B:APPA.0000006548.01625.72>

Coelho FMG (2005) O café num outro retrato do Brasil rural: o lugar da agricultura familiar. Informe Agropecuário, Belo Horizonte, Edição especial 26: 9-16.

Conab (2020). Companhia Nacional de Abastecimento. Acompanhamento da safra brasileira de café, Safra 2020, primeiro levantamento, Brasília, 6: 1-62.

Conab (2021) Companhia Nacional de Abastecimento. Acompanhamento da safra brasileira de café, Safra 2021, segundo levantamento, Brasília, pp 1-63.

Croft BA, Blackwood JS, McMurtry JA (2004) Classifying life-style types of phytoseiid mites: diagnostic traits. Exp. Appl. Acarol. 33: 247-21 260. <https://doi.org/10.1023/B:APPA.0000038622.26584.82>

Da Silva RTL, Da Silva VL, Silva DE et al (2020) Bioecological aspects of mites associated with *Vitis vinifera* varieties in the state of Rio Grande Do Sul, Brazil. Syst. Appl. Acarol. 25: 1618-1642. <https://doi.org/10.11158/saa.25.9.8>

Flechtmann CH (1967) Os ácaros do cafeiro. Anais da Escola Superior de Agricultura Luiz de Queiroz 24: 91-95.

Jeppson LR, Keifer HH, Baker EW (1975) Mites Injurious to Economic Plants. Univ. Calif. Press, Berkeley, pp 614.

Khan S, Khan MA, Hanjra MA, Mu J (2009) Pathways to reduce the environmental footprints of water and energy inputs in food production. Food policy 34: 141-149. <https://doi.org/10.1016/j.foodpol.2008.11.002>

Kitajima EW, Chagas CM, Rodrigues JC (2003) *Brevipalpus*-transmitted plant virus and virus-like diseases: cytopathology and some recent cases. Exp. Appl. Acarol. 30: 135–160. <http://dx.doi.org/10.1023/B:APPA.0000006546.55305.e3>

Koptur S (1994) Floral and extrafloral nectars of Costa Rican Inga trees: a comparison of their constituents and composition. *Biotropica* 26: 276-284. <https://doi.org/10.2307/2388848>

Le Pelley RH (1973) Coffee insects. *Annu. Rev. Entomol.* 18: 121–142.

Letourneau DK, Armbrecht I, Rivera BS et al (2011) Does plant diversity benefit agroecosystems? A synthetic review. *Ecol Appl* 21: 9-21. <https://doi.org/10.1890/09-2026.1>

Marchetti MM (2007) Ácaros do cafeiro em Minas Gerais com chave de identificação. Dissertation, Universidade Federal de Viçosa.

Matiello JB (1987) Novas condições de ocorrência de mancha anular do cafeiro. In 14. Congresso Brasileiro de Pesquisas Cafeeiras 1-4 Dic 1987 Campinas, SP (Brasil). Ministério da Indústria e do Comércio, Rio de Janeiro, RJ (Brasil) Instituto Brasileiro do Café, Rio de Janeiro.

Melo Y, Córdula E, Machado SR, Alves M (2010) Morfologia de nectários em Leguminosae senso lato em áreas de caatinga no Brasil. *Acta Bot. Croat.* 24: 1034-1045.

McMurtry JA, Croft BA (1997) Life-styles of phytoseiid mites and their roles in biological control. *Annu Rev Entomol* 42: 291-321. <https://doi.org/10.1146/annurev.ento.42.1.291>

McMurtry JA, Moraes GJD, Sourassou NF (2013) Revision of the lifestyles of phytoseiid mites (Acari: Phytoseiidae) and implications for biological control strategies. *Syst. Appl. Acarol.* 18: 297-320. <https://doi.org/10.11158/saa.18.4.1>

Mineiro JLC, Sato ME (2019) Distribuição de *Brevipalpus papayensis* Baker e *Brevipalpus yothersi* baker (Acari: Tenuipalpidae) em cafeeiros no estado de São Paulo. X Simpósio de Pesquisa dos Cafés do Brasil.

Mineiro JLDC, Sato ME, Matioli AL, Raga A (2009). Ácaros plantícolas e edáficos em agroecossistema cafeiro, em Atibaia, estado de São paulo. Simpósio de Pesquisa dos Cafés do Brasil, Brasília

Mineiro JLDC, Sato ME, Raga A, Arthur V (2008) Population dynamics of phytophagous and predaceous mites on coffee in Brazil, with emphasis on *Brevipalpus phoenicis* (Acari: Tenuipalpidae). *Exp. Appl. Acarol.* 44: 277-291. <https://doi.org/10.1007/s10493-008-9149-7>

Mineiro JLDC, Sato ME, Raga A, Arthur V, Cangani KG, Barbosa FV (2006a) Diversidade de ácaros (Arachnida: Acari) em cinco cultivares de duas espécies de cafeeiros (*Coffea* spp.) em Garça, Estado de São Paulo. *Arquivos do Instituto Biológico*, 73: 333-341.

Mineiro JLDC, Sato ME, Raga A, Arthur V, Moraes GJD, Sarreta FDO, Carrijo A (2006b) Diversidade de ácaros (Arachnida: Acari) em *Coffea arabica* L. cv. Mundo Novo, nos municípios de Jeriquara e Garça, Estado de São Paulo. *Biota Neotrop.* 6. <https://doi.org/10.1590/S1676-06032006000200012>

Moraes GJ (1987) Importance of taxonomy in biological control. *Int. J. Trop. Insect Sci.* 8: 841-844. <https://doi.org/10.1017/S1742758400023031>

Morris KS, Mendez VE, Lovell ST, Olson M (2013) Conventional food plot management in an organic coffee cooperative: Explaining the paradox. *Agroecol. Sustain. Food Syst.* 37: 762-787. <https://doi.org/10.1080/21683565.2013.774303>

Navia D, Mendonça RS, Ferragut F, Miranda LC, Trincado RC, Michaux J, Navajas M (2013) Cryptic diversity in *Brevipalpus* mites (Tenuipalpidae). *Zool. Scr.* 42: 406–426. <https://doi.org/10.1111/zsc.12013>

Nunes MA, de Carvalho Mineiro JL, Rogerio LA, Ferreira LM, Tassi A, Novelli VM, Kitajima EW, Freitas-Astúa J (2018) First report of *Brevipalpus papayensis* as vector of Coffee ringspot virus and Citrus leprosis virus C. *Plant Dis* 102.5: 1046-1046. <https://doi.org/10.1094/PDIS-07-17-1000-PDN>

Pallini A (1991) Acarofauna e predação de ácaros fitófagos por ácaros predadores em cafeeiro (*Coffea arabica* L.) no Sul de Minas Gerais. Dissertation, Universidade Federal de Lavras.

- Pedlowski MA, Canela MC, Terra MAC, de Faria RMR (2012) Modes of pesticides utilization by Brazilian smallholders and their implications for human health and the environment. *Crop Prot.* 31:113-118. <https://doi.org/10.1016/j.cropro.2011.10.002>
- Peña JE, Bullock RC (1994) Effects of feeding of broad mite (Acari: Tarsonemidae) on vegetative plant growth. *Fla Entomol* 77: 180-184. <https://doi.org/10.2307/3495885>
- Perfecto I, Rice R, Greenberg R, Van Der Voolt M (1996) Shade coffee as refuge of biodiversity. *BioScience* 46: 589–608. <https://doi.org/10.2307/1312989>
- Perfecto I, Vandermeer J (2008) Biodiversity conservation in tropical agroecosystems: a new conservation paradigm. *Annals of the New York Academy of Sciences*, 1134 : 173-200. [10.1196/annals.1439.011](https://doi.org/10.1196/annals.1439.011)
- Ratnadass A, Fernandes P, Avelino J, Habib R (2012) Plant species diversity for sustainable management of crop pests and diseases in agroecosystems: a review. *Agron Sustain Dev* 32: 273-303. [10.1007/s13593-011-0022-4](https://doi.org/10.1007/s13593-011-0022-4)
- Reis PR (2005) Ácaro-vermelho. Cultivar grandes culturas. *Pelotas* 7:14–17.
- Reis PR, Souza JC (1986) Pragas do cafeeiro. In: Rena AB, Malavolta E, Rocha M, Yamada T. *Cultura do cafeeiro: Fatores que afetam a produtividade*. Piracicaba: Associação Brasileira para Pesquisa da Potássio e do Fosfato. 323-378 pp.
- Rezende MQ, Venzon M, dos Santos PS, Cardoso IM, Janssen A (2021) Extrafloral nectary-bearing leguminous trees enhance pest control and increase fruit weight in associated coffee plants. *Agric. Ecosyst. Environ.* 319: 107538. <https://doi.org/10.1016/j.agee.2021.107538>
- Rezende MQ, Venzon M, Perez AL, Cardoso IM, Janssen A (2014) Extrafloral nectaries of associated trees can enhance natural pest control. *Agric. Ecosyst. Environ.* 188: 198-203. <https://doi.org/10.1016/j.agee.2014.02.024>
- Rosado CM, de Araújo GJ, Pallini A, Venzon M (2021) Cover crop intercropping increases biological control in coffee crops. *Biol Control* 160: 104675. <https://doi.org/10.1016/j.biocontrol.2021.104675>

Sales EF, Méndez VE, Caporal FR, Faria JC (2013) Agroecological transition of conilon coffee (*Coffea canephora*) agroforestry systems in the State of Espírito Santo, Brazil. *Agroecol. Sustain. Food Syst.* 37: 405-429.
<https://doi.org/10.1080/10440046.2012.712633>

Souza HN, Cardoso IM, Fernandes JM et al (2010) Selection of native trees for intercropping with coffee in the Atlantic Rainforest biome. *Agrofor. Syst* 80: 1-16.
<https://doi.org/10.1007/s10457-010-9340-9>

Souza JC, Reis PR, Rigitano RLO (1998) Bicho-mineiro-do-cafeeiro: biologia, danos e manejo integrado, EPAMIG, Boletim Técnico 54, Belo Horizonte, 2:1-48.

Spongowski S, Reis PR, Zacarias MS (2005) Acarofauna da cafeicultura de cerrado em Patrocínio, Minas Gerais. Ciênc. agrotec. 29: 9-17. <https://doi.org/10.1590/S1413-1570542005000100001>

Teodoro AV, Klein AM, Tscharntke T (2009) Temporally mediated responses of the diversity of coffee mites to agroforestry management. *J. Appl. Entomol.* 133: 659-665.
<https://doi.org/10.1111/j.1439-0418.2009.01422.x>

Thrupp LA (1990) Inappropriate incentives for pesticide use: Agricultural credit requirements in developing countries. *Agric Human Values* 7: 62-69.

Tixier MS (2018) Predatory mites (Acari: Phytoseiidae) in agro-ecosystems and conservation biological control: a review and explorative approach for forecasting plant-predatory mite interactions and mite dispersal. *Front. Ecol. Evol.* 6.
<https://doi.org/10.3389/fevo.2018.00192>

Chapter 1

Intercropped plants as a reservoir of predatory mites in coffee crop

Júlia Jantsch Ferla¹, Pedro Hermano Marques Gonçalves Nascimento¹, Milena Kalile¹, Shauanne Dias Pancieri¹, André Costa Cardoso¹, Gustavo Júnior de Araújo¹, Elem Fialho Martins¹, Madelaine Venzon², Noeli Juarez Ferla³, Angelo Pallini¹

¹ Department of Entomology, Federal University of Viçosa, Minas Gerais, Brazil

²Agriculture and Livestock Research Enterprise of Minas Gerais (EPAMIG), Viçosa, MG, Brazil.

³ Laboratório de Acarologia, Universidade do Vale do Taquari - Univates, Lajeado, Brazil.

Abstract

Conservation biological control of pest mites can be an alternative to pesticides in agroecosystems. The use of conservation biological control practices could be done with a variety of integrated strategies using the local biodiversity. Pollen and extrafloral nectar are important food source to some mites, making the intercropping systems with plants that provide this diet a suitable strategy to attract natural enemies of pests. We investigate if the communities of predators and phytophagous mites on coffee crops change along a distance gradient from selected Intercropped Plants Species (IPS) that provide nectar and pollen: *Inga edulis* Mart., *Senna macranthera* (Dc. ex collad.) H.S. Irwin & Barnaby and *Varronia curassavica* Jacq. We also assessed the composition of these mites on IPS. To do so, we recorded the mites species on coffee along transects of 16 m extending from the IPS and we sampled the mites on IPS. The community of predators and phytophagous mites on coffee was not modified by the distance from IPS, but IPS harbor several mite species of mites. Furthermore, it was found that the composition of predators and phytophagous mites among the IPS was different. The

findings suggest that the intercropped plants with coffee can attract and serve as a reservoir of predatory mites.

Keywords: Conservative biological control, Agroecosystem diversification, Ecosystem service, Phytophagous mites.

Introduction

Brazil is the main coffee producer and the largest exporter in the world, and the species most cultivated in the country is *Coffea arabica* L. (Rubiaceae) (Conab 2021). Minas Gerais is the state with the largest crop area, corresponding to 71.7% of the area occupied with this species nationwide (Conab 2021). Pests and diseases occurring in this culture can cause economic yield and quality losses (Avelino et al. 2018). The most frequent strategy in controlling them is the use of chemical inputs, such as: fungicides, insecticides and acaricides (Spongoski et al. 2005; Aparecido et al. 2020). However, the impact of these agrochemicals does not only affect the target pest species, but also many others that can play important roles in maintaining the ecosystem due to the services they provide, such as pollinators and natural enemies (Tasei 2002; Desneux et al. 2007; Giglio et al. 2017). In addition to the impact on biodiversity, the continued use of this practice also affects the quality of human life, as ecosystem services are lost, increasing the costs of food production (Perfecto et al. 1996; Perfecto and Vandermeer 2008; Khan et al. 2009; Pedlowski et al. 2012; Sales et al. 2013).

Conservation biological control is the enhancement of natural enemies populations in the agro-ecosystem and can be an alternative to conventional practices to control pests in the crop (Letourneau et al. 2011; Ratnadass et al. 2012; Tixier 2018). This management could be done with a variety of integrated strategies using the local biodiversity (Zehnder et al. 2007; Venzon et al. 2011). In management strategies, such as the intercropped systems, which aim to reduce the pest population throughout the increasing of natural enemy presence, it is important that introduced plants are effective in attracting and favors enemy target but not pests in crops (Venzon and Sujii 2009; Begg et al. 2017). Extrafloral nectar and pollen are important sources of energy that can attract natural enemies, such as insects and mites and also pollinators (i.e. wasps, ants, mites, bees, chrysopids), making the intercropping systems a suitable

strategy of conservative biological control in crops, such as coffee (Pemberton and Vandenberg 1993; van Rijn and Tanigoshi 1999; Wäckers et al. 2005; Rezende et al. 2014, 2021; Botti et al. 2021; Martins et al. 2021; Rosado et al. 2021).

Some predatory mites, such as phytoseiid from the genus *Amblyseius*, *Euseius*, *Neoseiulus*, *Galendromus* and *Iphiseiodes* for instance, can feed on phytophagous mites or some plant sources as pollen and nectar (McMurtry and Croft 1997; Croft 2004; McMurtry et al. 2013). There are some studies with predatory mites associated with extrafloral nectar (Pemberton 1993; Pemberton and Vandenberg 1993; Van Rijn and Tanigoshi 1999), pollen (McMurtry 1977; Villanueva and Childers 2004; Rodríguez-Cruz et al. 2013) and spontaneous plants intercropped with the main crop (Rosado et al. 2021). However, little is known about the effect of intercropped plants with nectaries and pollen on herbivores and predatory mites in coffee crops (Teodoro et al. 2009; Peixoto et al. 2017; Rosado et al. 2021).

Coffee crops dwell mites with different food habits, such as predators, fungivores, micophagous, phytophagous and other species that does not have a well-known food source reported (Pallini 1991; Reis et al. 2000; Spongoski et al. 2005; Marchetti 2007; Mineiro et al. 2008). Some of the most common predatory mites found on these plants are from the families Phytoseiidae, Ascidae, Stigmaeidae, Cunaxidae, Cheyletidae and Anystidae (Pallini 1991; Spongoski et al. 2005; Mineiro et al. 2006a; Marchetti 2007; Mineiro et al. 2008; Mineiro et al. 2009; Silva et al. 2010; Berton et al. 2019), being phytoseiid mites one of the most known and studied (McMurtry and Croft 1997). The phytophagous mites most economically important in this culture are *Oligonychus ilicis* (McGregor) (Tetranychidae), causing significant economic losses due to defoliation, premature leaf drop and reduction in plant photosynthesis (Reis and Souza 1986; Reis 2005) and *Brevipalpus phoenicis* (Geijskes) (Tenuipalpidae), that can transmit Coffee Ringspot Virus, causing defoliation of the plants (Chagas 1973; Reis and Zacarias 2007). This species used to comprise several cryptic species (Navia et al. 2013; Beard et al. 2015a). Beard et al. (2015a) divided it into a species complex of eight species. So, before this division, *B. phoenicis* were recorded on this crop, and after this there are records of *B. yothersi* Baker and *B. papayensis* Baker, and both species belong to *B. phoenicis* species complex and can transmit Coffee ringspot virus (Spongoski et al. 2005; Mineiro et al. 2006a, 2006b; Marchetti 2007; Mineiro et al.

2008, 2009; Nunes et al. 2018; Mineiro and Sato, 2019). Additionally, *Polyphagotarsonemus latus* (Banks) (Tarsonemidae) is also a pest on coffee, that damages the leaves, curling it downward (Amaral 1951; Flechtmann 1967; Chagas 1973; Jeppson et al. 1975; Matiello 1987; Peña and Bullock 1994).

Increasing plant diversity enhances the coffee productivity and the soil quality, attracts natural enemies and levels up predation and parasitism in this agroecosystem (Rezende et al. 2014; de Souza et al. 2017; Androcioli et al. 2018; Rezende et al. 2021). Therefore, our aim in this work was to evaluate if the communities of predators and phytophagous mites present on coffee crops change if their distance from the selected intercropped plants increase. For this, we used an established coffee system intercropped with providing nectar and pollen plants and evaluated whether the insertion of each intercropped plant modifies the abundance, richness and composition of predators and phytophagous mites. Our hypothesis is that coffee plants closer to the intercropped plants will have more predators, and consequently, less herbivores. Additionally, we evaluated the composition and species of predatory and phytophagous mites these plants can host to assess the role of these plants as a conservative biological control strategy.

Material and methods

Study area

The experiment was carried out at the EPAMIG Experimental Farm, in Patrocínio county ($18^{\circ}59'52.0"S$ $46^{\circ}58'59.8"W$), Minas Gerais state, Brazil, in Cerrado biome. The region is located at the mesoregion of Triângulo Mineiro and Alto Paranaíba (IBGE 2010). The sample area was characterized by a diversified system (1080 m^2), with 3 replicas in random blocks on 3 different places at the farm, away from at least 200m from each other. The diversified system is surrounded by two lines of three Intercropped Plant Species (IPS): two *Inga edulis* Mart. (Fabaceae) trees, one *Senna macranthera* (Dc. ex collad.) H.S. Irwin & Barnaby (Fabaceae) tree and six shrubs of *Varronia curassavica* Jacq. (Cordiaceae) in each line (Figure 1). *Inga edulis* and *S. macranthera* have extrafloral nectaries and *V. curassavica* produces flowers during the whole year (Koptur 1994; Melo et al. 2010; Brandão et al. 2015). No pesticides were applied in the study area. Fertilization was made with chemical fertilizers and the spontaneous plant growth was controlled by a brush cutter.

Data collection

Coffee plants were sampled along the lines of coffee plantation by detaching four leaves distally from the third leaf pair of two different branches. Two branches in the middle third of each bush facing both sides of the coffee lines were sampled (adapted Souza et al. 1998). There was a total of four plants, every 4 meters in a coffee line (4; 8; 12 and 16 m), with 16m extending from each IPS (Figure 1). One line in front of each IPS was sampled. It was a total of 192 leaves/area, totaling 576 leaves. As it was a huge amount of material and the mite species repeated itself, the fourth and fifth sampling were made the same way but instead of four leaves per plant, just 25% of the material were sampled, which means one leaf per plant. It was a total of 48 leaves/area, totaling 144 leaves (adapted Pallini 1991). To sample the IPS, five leaves of each plant species were detached randomly per line, with 10 leaves per plant species/area, totaling 90 leaves. The material was collected from June/2020 to Feb/2021, summing up 5 samplings in dry and rainy seasons.

The samples were placed separately in paper bags and kept at low temperature with Gelo-X® inside a cooler box until arrive to the laboratory of Acarology at the Department of Entomology from the Federal University of Viçosa, Viçosa, where it was kept in a refrigerator at approximately 10°C for a period up to 10 days. The material was examined with a stereoscopic microscope and all mites were mounted on glass slides in Hoyer's medium for identification (Krantz and Walter 2009). Morphological identification was made with a microscope with phase contrast and dichotomous key (André 1980; Baker and Tuttle 1994; Chant and McMurtry 1994, 2007; Beard et al. 2015a, 2015b, 2015c; Fan et al. 2016; Paktinat-Saeij et al. 2016).

To maintain a pattern, as the collections of coffee plants were made with different amounts of leaves, 25% of each slide from first to third collections was identified. To do so, the cover slide was divided into four identical quadrants, each one corresponding to 25% of the slide. The identified quadrant was raffled. If in the raffled quadrant there were no mites, the identified was the anterior one. In the fourth and fifth collections all the slide were identified.

To analyze abundance, females, male and immature mites were included but to richness, only females were included. When assessing Eriophyoidea mites, some individuals were mounted on microscope slides with modified Berlese's medium (Amrine and Manson 1996) for morphotyping and some individuals were preserved in

70% ethanol for posterior identification. To measure the abundance, all Eriophyoidea were counted in the first sample. To the second up to the fifth sample we did the mean number of mites per leaf. Hence, all individuals of the same morphospecies were counted in a 1 cm² quadrant in the abaxial surface of each leaf, in the region near to petiole and midrib. The leaves were measured and the number of specimens per leaf was estimated. Only to this superfamily, all specimens were included in richness.

All specimens were deposited in the mite reference collection of the Laboratory of Acarology in the Department of Entomology at Federal University of Viçosa, state of Minas Gerais, Brazil.

Statistical analyses

We used Generalized linear models (GLM) to test if the insertion of plants modifies the abundance of different groups of mites along the coffee line in front of the IPS. Since we had counting data, our model was fitted the family error Poisson. When we detected overdispersion in the residuals of the model, the Negative Binomial error was used. To determine the best-model we used Akaike Information Criterion corrected (AICc) comparing Akaike Information Criterion corrected (AICc) values, the difference in AICc between the model and the model with the smallest AICc (ΔAICc) and weights of each model (AICcWt) for values of the following sub-models: (1) Model 1: response variable ~ family * distance * plant; (2) Model 2: response variable ~ family * distance + plant; (3) Model 3: response variable ~ family + distance * plant; (4) Model: response variable ~ family + distance + plant, (5) Null model: response variable ~ 1. We obtained the best-model selecting that one with the lowest AICc throughout the function aictab from “AICcmodavg package” version 3.2-1 (Mazerolle 2020). To compare the differences in abundance and richness among families we performed a post-hoc analysis using the glht function from “multcomp package” version 1.4-17 (Hothorn et al. 2021). To test the variation in composition of predatory and phytophagous mites over distance and among plants, we used Permutational analysis of multivariate dispersions (PERMDISP) and Permutational multivariate analysis of variance (PERMANOVA) using the function betadisper and adonis, respectively, both from package ‘vegan’ version 2.4-2 (Oksanen et al. 2017). When we found differences in composition among treatments, we performed pairwise comparison using pairwise adonis function with Bonferroni correction (Arbizu 2019). All analyses were performed in R version 3.5.1 (R Core Team 2018)

Results

Abundance and richness on coffee and IPS

In total, we collected 8259 specimens belonging to 8 families and 28 species. Of the 28 species, 16 were predators (2 Iolinidae, 12 Phytoseiidae and 2 Stigmeidae) and 12 were phytophagous (4 Eriophyidae, 1 Diptilomiopidae, 1 Tarsonemidae, 1 Tenuipalpidae and 5 Tetranychidae). On coffee, Tenuipalpidae was the most abundant family ($n= 656$) but only one species was reported, followed by Tetranychidae ($n=96$) with 2 species and Phytoseiidae ($n=67$) with 4 species. On IPS, Eriophyidae was the most abundant family ($n= 6252$) with 4 species, followed by Tenuipalpidae ($n=257$) with 1 species and Phytoseiidae ($n=282$) with 11 species (Table 1, Table 2).

Diversity and composition on coffee crops

The best model that described the mite abundance was *families + distance + plant* ($AICc = 698.14$; and $AICcWt = 0.84$) (Tab. S1). We found statistical differences in Family abundance ($X^2 = 626.83$, $df = 4$, $p > 0.001$). No difference was found to distance ($X^2 = 5.80$, $df = 3$, $p = 0.12$) and plant ($X^2 = 1.13$, $df = 2$, $p = 0.56$, $R^2=0.59$).

In the coffee system studied, Tenuipalpidae was the most abundant family and Stigmeidae was the least one. We also found differences in Stigmeidae vs. Iolinidae, Phytoseiidae, Tenuipalpidae and Tetranychidae, as well to Tenuipalpidae vs. Iolinidae, Phytoseiidae and Tetranychidae, and Tetranychidae vs. Iolinidae (Figure 2, Table 3).

The best model that described the mite richness was *families + distance + plant* ($AICc = 331.16$; and $AICcWt = 1.00$) (Tab. S1). We found statistical differences in Family richness ($X^2 = 39.537$, $df = 4$, $p > 0.001$). No difference was found to distance ($X^2 = 0.899$, $df = 3$, $p = 0.82$) and plant ($X^2 = 0.14$, $df = 2$, $p = 0.93$, $R^2=0.39$).

The richness was similar in all families, but it differs from Stigmeidae vs. Iolinidae, Phytoseiidae, Tenuipalpidae and Tetranychidae (Figure 3, Table 4).

We did not find variation in composition between mite communities along distance in coffee plants (PERMDISP: $F = 0.07$ $p = 0.93$; PERMANOVA; $R^2 = 0.12$, $p = 0.63$; Figure 4). But the composition of phytophagous on IPS was different (PERMDISP: $F= 0.21$, $p = 0.80$; PERMANOVA: $R^2 = 0.22$, $p < 0.05$, Figure 5A). *Inga edulis* was different from *S. macranthera* ($R^2 = 0.32$, $p < 0.05$) and *V. curassavica* (R^2

=0.32, P<0.05), but we did not find difference between *V. curassavica* and *S. macranthera* ($R^2= 0.09$, $p = 0.36$).

The composition of predators on IPS was also different (PERMDISP: $F= 0.18$, $p =0.83$; PERMANOVA: $R^2 = 0.33$, $p < 0.001$, Figure 5B). *Inga edulis* were different from *V. curassavica* ($R^2 =0.32$, P<0.05) and *S. macranthera* ($R^2 =0.32$, P<0.05), but *V. curassavica* and *S. macranthera* was not different ($R^2 =0.09$, $P=0.37$).

The predatory mite species commons in all IPS were *Pseudopronematus nadirae* Silva, Da-Costa & Ferla, *Euseius citrifolius* (Denmark & Muma), *E. concordis* (Chant), *E. sibelius* (De Leon), *Typhlodromalus aripo* DeLeon and *Agistemus floridanus* Gonzalez. In general, *I. edulis* houses more predator species than the other IPS, with *Amblyseius* sp., *E. alatus* DeLeon, *Iphiseiodes zuluagai* Denmark & Muma and *Agistemus brasiliensis* Matioli, Ueckermann & Oliveira occurring only on this plant species. *Varronia curassavica* have only one exclusive species (*Neoseiulus tunus* (De Leon)) and *S. macranthera* did not have any exclusive species.

Brevipalpus yothersi and *Atrichoprocus uncinatus* Flechtmann were the only phytophagous mite species common in all IPS. Eriophyidae sp1., *O. ilicis* and *Tetranychus* sp. are the species exclusive of *I. edulis*. Eriophyidae sp2 and sp3, Diptilomiopidae sp. and *P. latus* were the species that occurs only on *V. curassavica* and *Aculus* sp. and *Mononychellus planki* (McGregor) are the only two species exclusive from *S. macranthera*. The number of Eriophyidae on *I. edulis* is high (4885), despite having only one species, while *V. curassavica* and *S. macranthera* have different species, and in a much lower number. It is important to notice that despite being a pest on coffee, *P. latus* only occurs on *V. curassavica* with no records on coffee.

Discussion

The data presented here shows that the plants intercropped are a reservoir of natural enemies as it harbors several species of predators. However, it does not modify the community of predators and phytophagous mites in coffee plants along the distance.

Diversified coffee production systems attract predators, mostly wasps and parasitoids, increases biological control, improves soil quality and decreases the spontaneous plants (Silva et al. 2013; Rezende et al. 2014; De Souza et al. 2017; Peixoto et al. 2017; Androcioli et al. 2018; Rezende et al. 2021; Rosado et al. 2021). Studies with *Inga* spp. demonstrate the positive effects of this plant intercropped with coffee increasing natural control of coffee berry borers and coffee leaf miners, important coffee pests, by attracting wasps and parasitoids due to the provision of nectar and consequently enhancing the coffee production (Rezende et al. 2014, 2021). So, in this work we can see that not just *I. edulis* so as *V. curassavica* and *S. macranthera* houses predators. All predatory mites found on coffee were also found on IPS (except for *A. aff impressus* that have only one individual) meaning that coffee predators are benefiting from IPS, but what is the exactly source of attraction and benefits are not clear. Besides providing additional food sources such as pollen and nectar, the intercropped plants also can provide shelter, favorable microclimate and a habitat where preys are present (Landis et al. 2000).

The composition of phytophagous and predaceous mites were different in *I. edulis* and similar in *V. curassavica* and *S. macranthera*. *Inga* plants harbored much more predators than other IPS and did not harbored *P. latus* and just few *O. ilicis* (n=1), two herbivore pests of economic impact on coffee crops. *Senna macranthera* doesn't host any of these pest species and also harbors some predators. This is an important finding because these plants have the potential of harboring different natural enemies and do not attract these coffee pest species.

The natural enemies found in this work belong to the families Stigmaeidae, Iolinidae and Phytoseiidae, being the last one with the most number of species. Most of the species found here are known as feeding and reproducing on a wide range of prey and in the case of Phytoseiidae, pollen and nectar can constitute an important part of their diet (Hessein and Perring 1988; Thistlewood et al. 1996; McMurtry and Croft 1997; Abou-Awad et al. 1999; Horn et al. 2011; McMurtry et al. 2013; Van de Velde et al. 2021). This is an important predatory family, extensively studied and used for biological control of pests (Gerson et al. 2003). Species of the genus *Amblyseius*, *Neoseiulus*, *Galendromus* and *Typhlodromalus* have records of feeding on pollen, Eriophyidae, Tarsonemidae, Tetranychidae, Tydeoidea and other families (McMurtry

and Croft 1997; McMurtry et al. 2013). Besides that, *Euseius* and *Iphiseiodes* are pollen feeding generalists that prey on a range of mite families, being *B. phoenicis* one of them (McMurtry et al. 2013; Reis et al. 1998; Reis et al. 2003; Yamamoto and Gravena 1996). At the time of studies cited above, the *B. phoenicis* were not divided into a species complex yet (Beard et al. 2015a) and all the species of these complex were called just *B. phoenicis*, so we cannot know for sure which species of these complex this works refers. Additionally, *I. zuluagai* can feed on sugary substances, being able to be reared on a range of alternative food sources (Yamamoto and Gravena 1996; Albuquerque and Moraes 2008).

The feeding habits of Tydeoidea are diverse (André and Fain 2000) and Iolinidae are associated with several phytophagous species, preying Eriophyidae, Tenuipalpidae and Tetranychidae (Hessein and Perring 1988; Abou-Awad et al. 1999; Horn et al. 2011; Van de Velde et al. 2021). Other mites that feed on Eriophyoidea are Stigmaeidae, which also can feed on Tetranychidae (White 1976; Thistlewood et al. 1996). In this work there were a great amount of Eriophyidae and other phytophagous species belonging to Diptilomiopidae, Tenuipalpidae, Tarsonemidae and Tetranychidae. Knowing that the pests on coffee vary in quantity during the year, probably the phytophagous on IPS can be preyed and serve as alternative food by the predators recorded here, mainly when the pests on coffee are in low population (Pallini 1991; Reis and Zacarias 2007; Franco et al. 2008). Besides, the IPS stimulate diversity of natural enemies of other insect pests in this crop (Landis et al. 2005; Rezende et al. 2014, 2021).

Thus, these results suggest that IPS attract predator and herbivore mites that are not pests on coffee. These herbivores could be an alternative prey to increase the population of predators on these plants that could later migrate to coffee plants and control their pest mites. Once this happens, it could enhance conservation biological control in the area. Understanding how the dynamics of predators and prey in this system would facilitate management practices and favor the establishment of a sustainable pest mite control on coffee crops.

Acknowledgments

The authors are grateful to Professor Enrico de Lillo, Marcello De Giosa, Tairis Da-Costa and Gabriel Lima Bizarro for taxonomic discussion and help. We also thank the colleagues from the Laboratory of Acarology of the Federal University of Viçosa for assistance with practical work, suggestions and discussions. This study was financed in part by the Coordination for the Improvement of Higher Education Personnel (CAPES) finance code 001, Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Consórcio Brasileiro de Pesquisa e Desenvolvimento do Café (CBP & D-Café) and Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG).

References

- Abou-Awad BA, El-Sawaf BM, Abdel-Kader AA (1999) Life history and life table of *Pronematus ubiquitus* (McGregor) as a predator of eriophyoid mites in Egypt (Acari: Tydeidae). *Acarologia* 40: 29-32.
- Albuquerque FAD, Moraes GJD (2008) Perspectivas para a criação massal de *Iphiseiodes zuluagai* Denmark & Muma (Acari: Phytoseiidae). *Neotrop Entomol* 37: 328-333. <https://doi.org/10.1590/S1519-566X2008000300013>
- Amaral JF (1951) A infestação de ácaros nos cafezais. *Biol* 17 pp 130.
- Amrine JWJ, Manson DCM (1996) Preparation, mounting and descriptive study of Eriophyoid mites. In Lindquist EE, Sabelis MW, Bruun J (eds): *Eriophyoid Mites – Their Biology, Natural Enemies and Control, World Crop Pests*, Elsevier Science Publishers, Amsterdam, pp 383–396.
- André HM (1980) A generic revision of the family Tydeidae (Acari: Actinedida). IV. Generic descriptions, keys and conclusions. *Annales de la Societe royale Zoologique de Belgie* 116: 103 - 168
- André HM, Fain A (2000) Phylogeny, ontogeny and adaptive radiation in the superfamily Tydeoidea (Acari: Actinedida), with a reappraisal of morphological characters. *Zool J Linn Soc* 130: 405-448. <https://doi.org/10.1111/j.1096-3642.2000.tb01636.x>

Androcioli HG, Hoshino AT, Menezes Júnior ADO, Morais H, Bianco R, Caramori PH (2018) Coffee leaf miner incidence and its predation by wasp in coffee intercropped with rubber trees. *Coffee Sci* 13:389 – 400. <http://dx.doi.org/10.25186/cs.v13i3.1487>

Aparecido LEO, Rolim GS, Moraes JRSC, Costa CTS, de Souza PS (2020) Machine learning algorithms for forecasting the incidence of *Coffea arabica* pests and diseases. *Int. J. Biometeorol.* 64: 671–688. <https://doi.org/10.1007/s00484-019-01856-1>

Arbizu MP (2019) pairwiseAdonis: pairwise multilevel comparison using adonis.

Avelino J, Allinne C, Cerda R, Willocquet L, Savary S (2018) Multiple-disease system in coffee: From crop loss assessment to sustainable management. *Annu. Rev. Phytopathol.* 56: 611-635. <https://doi.org/10.1146/annurev-phyto-080417-050117>

Baker EW, Tuttle DM (1994) A guide to the spider mites (Tetranychidae) of the United States. Indira Publishing House, United States of America pp 347.

Beard JJ, Ochoa R, Braswell WE, Bauchan GR (2015a) *Brevipalpus phoenicis* (Geijskes) species complex (Acari: Tenuipalpidae) --a closer look. *Zootaxa* 3944: 1-67. <http://dx.doi.org/10.11646/zootaxa.3944.1.1>

Beard JJ, Ochoa R, Bauchan GR, Trice MD, Redford AJ, Walters TW, Mitter C (2015b) Lucid key: *Brevipalpus* of the world. Lucid key: *Brevipalpus* of the world.

Beard JJ, Ochoa R, Bauchan GR, Trice MD, Redford AJ, Walters TW, Mitter C (2015c) Lucid key: Flat mite genera of the world. Lucid key: Flat mite genera of the world.

Begg GS, Cook SM, Dye R, Ferrante M, Franck P, Lavigne C, Lövei GL, Mansion-Vaque A, Pell JK, Petit S, Quesada N, Ricci B, Wratten SD, Birch ANE (2017) A functional overview of conservation biological control. *Crop Prot* 97:145–158. <https://doi.org/10.1016/j.cropro.2016.11.008>

Berton LHC, Mineiro JLC, Sato ME, Joaquim ADAF, Raga A (2019) Mite fauna of a coffee agroecosystem (*Coffea arabica* L.) in the municipality of Monte Alegre do Sul, São Paulo State, Brazil. Part I. *Acarologia* 59: 542-550. [10.24349/acarologia/20194352](https://doi.org/10.24349/acarologia/20194352)

Botti JMC (2021) Role of green lacewing and ants of coffee berry borer predation. PhD thesis, Universidade Federal de Viçosa.

Botti JMC, Martins EF, Franzin ML, Venzon M (2021) Predation of Coffee Berry Borer by a Green Lacewing. *Neotrop Entomol* 1-4. <https://doi.org/10.1007/s13744-021-00884-0>

Brandão DS, Mendes ADR, Santos RR, Rocha SMG, Leite GLD, Martins ER (2015) Biologia floral e sistema reprodutivo da erva-baleeira (*Varronia curassavica* Jacq.). *Rev. Bras. Pl. Med.* 17: 562-569. [10.1590/1983-084X/14_011](https://doi.org/10.1590/1983-084X/14_011)

Chagas CM (1973) Associação do ácaro *Brevipalpus phoenicis* (Geijskes) à mancha anular do cafeeiro. *Biol* 39: 229-232.

Chant DA, McMurtry JA (1994) A review of the subfamilies Phytoseiinae and Typhlodrominae (Acari: Phytoseiidae). *Int J Acarol* 20:223–310.

Chant DA, McMurtry JA (2007) Illustrated keys and diagnoses for the genera and subgenera of the Phytoseiidae of the world (Acari: Mesostigmata). Indira Publishing House, West Bloomfield pp 220.

Conab (2021) Companhia Nacional de Abastecimento. Acompanhamento da safra brasileira de café, Safra 2021, segundo levantamento, Brasília, pp 1-63.

Croft BA, Blackwood JS, McMurtry JA (2004) Classifying life-style types of phytoseiid mites: diagnostic traits. *Exp Appl Acarol* 33: 247-260. <https://doi.org/10.1023/B:APPA.0000038622.26584.82>

Desneux N, Decourtey A, Delpuech JM (2007) The sublethal effects of pesticides on beneficial arthropods. *Annu. Rev. Entomol.* 52: 81-106. <https://doi.org/10.1146/annurev.ento.52.110405.091440>

De Souza GSD, Alves DI, Dan ML, Lima JS, Fonseca ALCCD, Araújo JBS, Guimarães LADOP (2017) Soil physico-hydraulic properties under organic conilon coffee intercropped with tree and fruit species. *Pesqui Agropecu Bras* 52: 539-547. <https://doi.org/10.1590/S0100-204X2017000700008>

Fan QH, Flechtmann CH, Moraes GJ (2016) Annotated catalogue of Stigmeidae (Acari: Prostigmata), with a pictorial key to genera. Zootaxa 4176: 1-199. 10.11646/ZOOTAXA.4176.1.1

Flechtmann CH (1967) Os ácaros do cafeiro. Anais da Escola Superior de Agricultura Luiz de Queiroz 24: 91-95.

Franco RA, Reis PR, Zacarias MS, Altoé BF, Neto MP (2008) Dinâmica populacional de *Oligonychus ilicis* (McGregor, 1917) (Acari: Tetranychidae) em cafeiro e de fitoseídeos associados a ele. Coffee Sci 3: 38-46.

Gerson U, Smiley RL, Ochoa R (2003) Mites (Acari) for pest control. Blackwell Science, Oxford, pp 539.

Giglio A, Cavaliere F, Julianini PG, Mazzei A, Talarico F., Vommaro, M. L., & Brandmayr, P. (2017) Impact of agrochemicals on non-target species: *Calathus fuscipes* Goeze 1777 (Coleoptera: Carabidae) as model. Ecotoxicol Environ Saf 142: 522-529. <https://doi.org/10.1016/j.ecoenv.2017.04.056>

Hessein NA, Perring TM (1988) *Homeopronematus anconai* (Baker)(Acari: Tydeidae) predation on citrus flat mite, *Brevipalpus lewisi* McGregor (Acari: Tenuipalpidae). Int J Acarol 14: 89-90. <https://doi.org/10.1080/01647958808683791>

Horn TB, Johann L, Ferla NJ (2011) Ecological interactions between phytophagous and predaceous mites in citrus agroecosystems in Taquari Valley, Rio Grande do Sul, Brazil. Syst Appl Acarol 16: 133-144. <https://doi.org/10.11158/saa.16.2.2>

Hothorn T, Bretz F, Westfall P, Heiberger RM, Schuetzenmeister A, Scheibe S (2021) Simultaneous Inference in General Parametric Models Version 1.4-17.

IBGE (2010) Estado de Minas Gerais meso e microrregiões do IBGE. https://www.mg.gov.br/sites/default/files/paginas/arquivos/2016/ligminas_10_2_04_lis_tamesomicro.pdf. Accessed 27 July 2021.

Jeppson LR, Keifer HH, Baker EW (1975) Mites Injurious to Economic Plants. Univ. Calif. Press, Berkeley, pp 614.

Khan S, Khan MA, Hanjra MA, Mu J (2009) Pathways to reduce the environmental footprints of water and energy inputs in food production. *Food policy* 34: 141-149. <https://doi.org/10.1016/j.foodpol.2008.11.002>

Koptur S (1994). Floral and extrafloral nectars of Costa Rican Inga trees: a comparison of their constituents and composition. *Biotropica* 26: 276-284. <https://doi.org/10.2307/2388848>

Krantz GW, Walter DE (2009) A manual of acarology. Third Edition. USA, Texas Tech University Press, pp 807.

Landis DA, Menalled FD, Costamagna AC, Wilkinson TK (2005) Manipulating plant resources to enhance beneficial arthropods in agricultural landscapes. *Weed Sci* 53: 902–908. <https://doi.org/10.1614/WS-04-050R1.1>

Landis DA, Wratten SD, Gurr GM (2000) Habitat management to conserve natural enemies of arthropod pests in agriculture. *Annu Rev Entomol* 45: 175-201. <https://doi.org/10.1146/annurev.ento.45.1.175>

Letourneau DK, Armbrecht I, Rivera BS et al (2011) Does plant diversity benefit agroecosystems? A synthetic review. *Ecol Appl* 21: 9-21. <https://doi.org/10.1890/09-2026.1>

Marchetti MM (2007) Ácaros do cafeiro em Minas Gerais com chave de identificação. Dissertation, Universidade Federal de Viçosa.

Martins EF, Franzin ML, Perez AL, Schmidt JM, Venzon M (2021) Is *Ceraeochrysa cubana* a coffee leaf miner predator? *Biol Control* 160: 104691. <https://doi.org/10.1016/j.biocontrol.2021.104691>

Matiello JB (1987) Novas condições de ocorrência de mancha anular do cafeiro. In 14. Congresso Brasileiro de Pesquisas Cafeeiras 1-4 Dic 1987 Campinas, SP (Brasil). Ministério da Indústria e do Comércio, Rio de Janeiro, RJ (Brasil) Instituto Brasileiro do Café, Rio de Janeiro.

Mazerolle MJ (2020) Model selection and multimodel inference using the AICcmodavg package.

- McMurtry JA (1977) Some predaceous mites [Phytoseiidae] on citrus in the mediterranean region. *Entomophaga* 22: 19-30. <https://doi.org/10.1007/BF02372986>
- McMurtry JA, Croft BA (1997) Life-styles of phytoseiid mites and their roles in biological control. *Annu Rev Entomol* 42: 291-321. <https://doi.org/10.1146/annurev.ento.42.1.291>
- McMurtry JA, Moraes GJD, Sourassou NF (2013) Revision of the lifestyles of phytoseiid mites (Acari: Phytoseiidae) and implications for biological control strategies. *Syst. Appl. Acarol.* 18: 297-320. <https://doi.org/10.11158/saa.18.4.1>
- Melo Y, Córdula E, Machado SR, Alves M (2010) Morfologia de nectários em Leguminosae senso lato em áreas de caatinga no Brasil. *Acta Bot. Croat.* 24: 1034-1045.
- Mineiro JLDC, Sato ME (2019) Distribuição de *Brevipalpus papayensis* Baker e *Brevipalpus yothersi* baker (Acari: Tenuipalpidae) em cafeeiros no estado de São Paulo. X Simpósio de Pesquisa dos Cafés do Brasil.
- Mineiro JLDC, Sato ME, Matioli AL, Raga A (2009) Ácaros plantícolas e edáficos em agroecossistema cafeiro, em Atibaia, estado de São paulo. Simpósio de Pesquisa dos Cafés do Brasil, Brasília
- Mineiro JLDC, Sato ME, Raga A, Arthur V, Cangani KG, Barbosa FV (2006a) Diversidade de ácaros (Arachnida: Acari) em cinco cultivares de duas espécies de cafeeiros (*Coffea* spp.) em Garça, Estado de São Paulo. Arquivos do Instituto Biológico, 73: 333-341.
- Mineiro JLDC, Sato ME, Raga A, Arthur V, Moraes GJD, Sarreta FDO, Carrijo A (2006b) Diversidade de ácaros (Arachnida: Acari) em *Coffea arabica* L. cv. Mundo Novo, nos municípios de Jeriquara e Garça, Estado de São Paulo. Biota Neotrop. 6: 333-341. <https://doi.org/10.1590/S1676-06032006000200012>
- Mineiro JLDC, Sato ME, Raga A, Souza Filho, MFD, Spongoski S (2008) Incidência de ácaros em cafeiro cv. Catuaí Amarelo. *Bragantia*, 67: 197-201.

Navia D, Mendonça RS, Ferragut F, Miranda LC, Trincado RC, Michaux J, Navajas M (2013) Cryptic diversity in *Brevipalpus* mites (Tenuipalpidae). Zool. Scr. 42: 406–426. <https://doi.org/10.1111/zsc.12013>

Nunes MA, de Carvalho Mineiro JL, Rogerio LA, Ferreira LM, Tassi A, Novelli VM, Kitajima EW, Freitas-Astúa J (2018) First report of *Brevipalpus papayensis* as vector of Coffee ringspot virus and Citrus leprosis vírus C. Plant Dis 102.5: 1046-1046. <https://doi.org/10.1094/PDIS-07-17-1000-PDN>

Oksanen J, Kindt R, Legendre P, O'Hara B, Simpson GL, Solymos P, Stevens MHH, Wagner H (2017) R package version 2.4-2." Vegan: Community Ecology Package <https://CRAN.R-project.org/package=vegan>.

Paktinat-Saeij S, Bagheri M, Noronha ACDS (2016) A new species of *Agistemus* Summers (Acari: Trombidiformes: Stigmaeidae) from Brazil, with a key to the American species. Syst. Appl. Acarol. 21: 813-819. <https://doi.org/10.11158/saa.21.6.8>

Pallini A (1991) Acarofauna e predação de ácaros fitófagos por ácaros predadores em cafeeiro (*Coffea arabica* L.) no Sul de Minas Gerais. Dissertation, Universidade Federal de Lavras.

Pedlowski MA, Canela MC, Terra MAC, de Faria RMR (2012) Modes of pesticides utilization by Brazilian smallholders and their implications for human health and the environment. Crop Prot. 31:113-118. <https://doi.org/10.1016/j.cropro.2011.10.002>

Peixoto ML, Fernandes LG, Carvalho MAC, Oliveira MLD, Putti FF, Reis ARD (2017) Assessment of mite fauna in different coffee cropping systems in Brazil. Biocontrol Sci. Technol. 27: 424-432. <https://doi.org/10.1080/09583157.2017.1296931>

Pemberton RW (1993) Observations of extrafloral nectar feeding by predaceous and fungivorous mites. Proc. Entomol. Soc. Wash. 95: 642-643.

Pemberton RW, Vandenberg NJ (1993) Extrafloral nectar feeding by ladybird beetles (Coleoptera: Coccinellidae). Proc. Entomol. Soc. Wash. 95: 139-151.

Peña JE, Bullock RC (1994) Effects of feeding of broad mite (Acari: Tarsonemidae) on vegetative plant growth. Fla Entomol 77: 180-184. <https://doi.org/10.2307/3495885>

Perfecto I, Rice R, Greenberg R, Van Der Voolt M (1996) Shade coffee as refuge of biodiversity. BioScience 46: 589–608. <https://doi.org/10.2307/1312989>

Perfecto I, Vandermeer J (2008) Biodiversity conservation in tropical agroecosystems: a new conservation paradigm. Annals of the New York Academy of Sciences, 1134 : 173-200.

R Core Team (2018) A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna. <https://www.R-project.org/>

Ratnadass A, Fernandes P, Avelino J, Habib R (2012) Plant species diversity for sustainable management of crop pests and diseases in agroecosystems: a review. Agron Sustain Dev 32: 273-303. <https://doi.org/10.1007/s13593-011-0022-4>

Reis PR (2005) Ácaro-vermelho. Cultivar grandes culturas. Pelotas 7:14–17.

Reis PR, Chiavegato LG, Alves EB (1998) Biologia de *Iphiseiodes zuluagai* Denmark & Muma (Acari: Phytoseiidae). Anais da Sociedade Entomológica do Brasil 27: 185-191.

Reis PR, Chiavegato LG, Alve EB, Souza EO (2000) Ácaros da família Phytoseiidae associados aos citros no município de Lavras, sul de Minas Gerais. Annales da Sociedade Entomologica Brasileira 29: 95-104.

Reis PR, Souza JC (1986) Pragas do cafeeiro. In: Rena AB, Malavolta E, Rocha M, Yamada T. Cultura do cafeeiro: Fatores que afetam a produtividade. Piracicaba: Associação Brasileira para Pesquisa da Potássio e do Fosfato p. 323-378.

Reis PR, Souza EO, Teodoro AV, Pedro Neto M (2003) Effect of prey density on the functional and numerical responses of two species of predaceous mites (Acari: Phytoseiidae). Neotrop. Entomol. 32: 461-467. <https://doi.org/10.1590/S1519-566X2003000300013>

Reis PR, Zacarias MS (2007) Ácaros em cafeeiro. EPAMIG. Boletim Técnico 81: 1-76.

Rezende MQ, Venzon M, dos Santos PS, Cardoso IM, Janssen A (2021) Extrafloral nectary-bearing leguminous trees enhance pest control and increase fruit weight in

associated coffee plants. Agric. Ecosyst. Environ. 319: 107538.
<https://doi.org/10.1016/j.agee.2021.107538>

Rezende MQ, Venzon M, Perez AL, Cardoso IM, Janssen A (2014) Extrafloral nectaries of associated trees can enhance natural pest control. Agric. Ecosyst. Environ. 188: 198-203. <https://doi.org/10.1016/j.agee.2014.02.024>

Rodríguez-Cruz FA, Venzon M, Pinto CMF (2013) Performance of *Amblyseius herbicolus* on broad mites and on castor bean and sunnhemp pollen. Exp. Appl. Acarol. 60: 497-507. <https://doi.org/10.1007/s10493-013-9665-y>

Rosado CM, de Araújo GJ, Pallini A, Venzon M (2021) Cover crop intercropping increases biological control in coffee crops. Biol Control 160: 104675. <https://doi.org/10.1016/j.biocontrol.2021.104675>

Sales EF, Méndez VE, Caporal FR, Faria JC (2013) Agroecological transition of conilon coffee (*Coffea canephora*) agroforestry systems in the State of Espírito Santo, Brazil. Agroecol. Sustain. Food Syst. 37: 405-429. <https://doi.org/10.1080/10440046.2012.712633>

Souza JC, Reis PR, Rigitano RLDO (1998) Bicho-mineiro do cafeiro: biologia, danos e manejo integrado. Belo Horizonte, Brasil: EPAMIG. pp 48.

Silva VDC, Perdoná MJ, Soratto RP, Negrisoli E (2013) Ocorrência de plantas daninhas em cultivo consorciado de café e noqueira-macadâmia. Pesq. Agropec. Trop 43: 441-449.

Silva EA, Reis PR, Zacarias MS, Marafeli PP (2010) Fitoseídeos (Acari: Phytoseiidae) associados a cafezais e fragmentos florestais vizinhos. Cienc. e Agrotecnologia 34: 1146-1153. <https://doi.org/10.1590/S1413-70542010000500010>

Spongowski S, Reis PR, Zacarias MS (2005) Acarofauna da cafeicultura de cerrado em Patrocínio, Minas Gerais. Ciênc. agrotec. 29: 9-17. <https://doi.org/10.1590/S1413-70542005000100001>

Tasei JN (2002) Impact of agrochemicals on non-Apis bees. Honey bees: estimating the environmental impact of chemicals, Taylor and Francis, United States of America p. 101-131.

Teodoro AV, Klein AM, Tscharntke T (2009) Temporally mediated responses of the diversity of coffee mites to agroforestry management. *J. Appl. Entomol.* 133: 659-665. <https://doi.org/10.1111/j.1439-0418.2009.01422.x>

Thistlewood HMA, Clements DR, Harmsen R (1996) Stigmaeidae. *World Crop Pests*, Elsevier p. 457-470.

Tixier MS (2018) Predatory mites (Acari: Phytoseiidae) in agro-ecosystems and conservation biological control: a review and explorative approach for forecasting plant-predatory mite interactions and mite dispersal. *Front. Ecol. Evol.* 6. <https://doi.org/10.3389/fevo.2018.00192>

Van de Velde V, Duarte MV, Benavente A, Vangansbeke D, Wäckers F, De Clercq P (2021) Quest for the Allmitey: Potential of *Pronematus ubiquitus* (Acari: Iolinidae) as a biocontrol agent against *Tetranychus urticae* and *Tetranychus evansi* (Acari: Tetranychidae) on tomato (*Solanum lycopersicum* L.). *bioRxiv*. <https://doi.org/10.1101/2021.04.08.438973>

van Rijn, PC, Tanigoshi LK (1999) The contribution of extrafloral nectar to survival and reproduction of the predatory mite *Iphiseius degenerans* on *Ricinus communis*. *Ecology and Evolution of the Acari*, Springer, Dordrecht pp 405-417.

Venzon M, Amaral DSSL, Togni PHB, Rezende MQ, Perez AL (2011) Manejo de pragas na agricultura orgânica. *Tecnologias para produção orgânica*. EPAMIG Zona da Mata, Viçosa, pp 107-128.

Venzon M, Sujii ER (2009) Controle biológico conservativo. *Informe Agropecuário*, Belo Horizonte, 30: 16-123.

Villanueva RT, Childers CC (2004) Phytoseiidae increase with pollen deposition on citrus leaves. *Fla. Entomol.* 87: 609-611. <https://www.jstor.org/stable/3496456>

Wäckers FL, van Rijn PCJ, Bruin J (2005) Plant-provided food for carnivorous insects: a protective mutualism and its applications. Cambridge University Press, Amsterdam.

White NDG (1976) Some aspects of the biology of the predaceous mite *Zetzellia mali* (Ewing) (Acarina: Stigmaeidae) found in southern Ontario apple orchards. Dissertation, University of Guelph.

Yamamoto PT, Gravena S (1996) Influência da temperatura e fontes de alimento no desenvolvimento e oviposição de *Iphiseiodes zuluagai* Denmark & Muma (Acari: Phytoseiidae). An. Soc. Entomol. Brasil 25: 109-115.

Zehnder G, Gurr GM, Kühne S, Wade MR, Wratten SD, Wyss E (2007) Arthropod pest management in organic crops. Annu. Rev. Entomol. 52: 57-80.
<https://doi.org/10.1146/annurev.ento.52.110405.091>

Table 1: Abundances of mite species recorded on the cited plants between June 2020 and February 2021: **Vc** *Varronia curassavica*, **Sm** *Senna macranthera*, **Ie** *Inga edulis*, **Ca1** *Coffea arabica* line 1, **Ca2** *C. arabica* line 2, **Ca3** *C. arabica* line 3 and **Ca4** *C. arabica* line 4.

Mite family	Mite species	Plant species					
		Vc	Sm	Ie	Ca1	Ca2	Ca3
Eriophyidae	<i>Aculus</i> sp.						
Eriophyidae	<i>Eriophyidae</i> sp1						4885
Eriophyidae	<i>Eriophyidae</i> sp2				70		
Eriophyidae	<i>Eriophyidae</i> sp3				97		
Diplopiidae	<i>Diplopiidae</i> sp.				18		
Tarsonemidae	<i>Polyphagotarsonemus latus</i> (Banks)				65		
Tenuipalpidae	<i>Brevipalpus yothersi</i> Baker				15	144	77
Tetranychidae	<i>Atrichoprocus uncinatus</i> Flechtmann				12	7	12
Tetranychidae	<i>Mononychellus planki</i> (McGregor)				3		
Tetranychidae	<i>Oligonychus coffeeae</i> (Nietner)					7	4
Tetranychidae	<i>Oligonychus ilicis</i> (McGregor)				1	1	4
Tetranychidae	<i>Tetranychus</i> sp.				2		2
Phytophagous	<i>loloides</i>	<i>Pseudopronematus nadirae</i> Silva, Da-Costa & Ferla	41	61	1	5	7
	<i>loloides</i>	<i>Pausia</i> sp.		5	7	10	4
Predators							

Phytoseiidae	<i>Amblyseius aff. chiapensis</i>	1	2	
Phytoseiidae	<i>Amblyseius aff impressus</i>			1
Phytoseiidae	<i>Amblyseius sp.</i>		1	
Phytoseiidae	<i>Euseius alatus DeLeon</i>	4		
Phytoseiidae	<i>Euseius citrifolius</i> (Denmark & Muma)	5	6	2
Phytoseiidae	<i>Euseius concordis</i> (Chant)	1	1	2
Phytoseiidae	<i>Euseius sibellus</i> (De Leon)	1	11	1
Phytoseiidae	<i>Iphiseiodes zuluagai</i> Denmark & Muma	32		
Phytoseiidae	<i>Galendromus annectens</i> (De Leon)	4	2	
Phytoseiidae	<i>Neoseiulus tunus</i> (De Leon)	6		
Phytoseiidae	<i>Typhlodromalus aripo</i> DeLeon	4	4	1
Phytoseiidae	<i>Typhlodromips manglaeae</i> De Leon		1	4
Stigmaeidae	<i>Agistemus floridanus</i> Gonzalez	38	13	1
Stigmaeidae	<i>Agistemus brasiliensis</i> Matoli, Ueckermann & Oliveira		2	1

Table 2: Abundances of mite families recorded on the cited plants between June 2020 and February 2021: **Vc** *Varronia curassavica*, **Sm** *Senna macranthera*, **Ie** *Inga edulis*, **Ca1** *Coffea arabica* line 1, **Ca2** *C. arabica* line 2, **Ca3** *C. arabica* line 3 and **Ca4** *C. arabica* line 4.

	Mite family	Plant species						
		Vc	Sm	Ie	Ca1	Ca2	Ca3	Ca4
Phytophagous	Eriophyidae	167	1200	4885				
	Diptilomiopidae	18						
	Tarsonemidae	108						
Predators	Tenuipalpidae	15	158	84	130	208	171	147
	Tetranychidae	19	14	18	16	14	27	39
	Iolinidae	85	134	2	8	12	20	11
	Phytoseiidae	37	53	192	16	15	15	21
	Stigmaeidae	48	20	129	1		1	1

Table 3: Tukey contrasts between the abundance of different mite families recorded in coffee plants between June 2020 and February 2021

	Estimate	Std. Error	z value	Pr (> z)
Phytoseiidae - Iolinidae	0.2481	0.2155	1.151	0.7560
Stigmaeidae - Iolinidae	-2.8655	0.6038	-4.746	<0.001
Tenuipalpidae - Iolinidae	2.5244	0.1820	13.868	<0.001
Tetranychidae - Iolinidae	0.5989	0.2051	2.920	0.0242
Stigmaeida - Phytoseiidae	-3.1136	0.6003	-5.186	<0.001
Tenuipalpidae - Phytoseiidae	2.2763	0.1702	13.377	<0.001
Tetranychidae - Phytoseiidae	0.3508	0.1947	1.802	0.3406
Tenuipalpidae - Stigmaeidae	5.3899	0.5892	9.148	<0.001
Tetranychidae - Stigmaeidae	3.4644	0.5967	5.806	<0.001
Tetranychidae - Tenuipalpidae	-1.9255	0.1567	-12.286	<0.001

Table 4: Tukey contrasts between the richness of different mite families recorded in coffee plants between June 2020 and February 2021

	Estimate	Std. Error	z value	Pr (> z)
Phytoseiidae - Iolinidae	0.39087	0.26998	1.448	0.57553
Stigmeidae - Iolinidae	-2.03688	0.61374	-3.319	0.00697
Tenuipalpidae - Iolinidae	0.44802	0.26694	1.678	0.42518
Tetranychidae - Iolinidae	0.26570	0.27715	0.959	0.86363
Stigmeida - Phytoseiidae	-2.42775	0.60217	-4.032	<0.001
Tenuipalpidae- Phytoseiidae	0.05716	0.23914	0.239	0.99921
Tetranychidae - Phytoseiidae	-0.12516	0.25049	-0.500	0.98619
Tenuipalpidae - Stigmeidae	2.48491	0.60081	4.136	<0.001
Tetranychidae - Stigmeidae	2.30259	0.60542	3.803	0.00110
Tetranychidae - Tenuipalpidae	-0.18232	0.24721	-0.738	0.94310

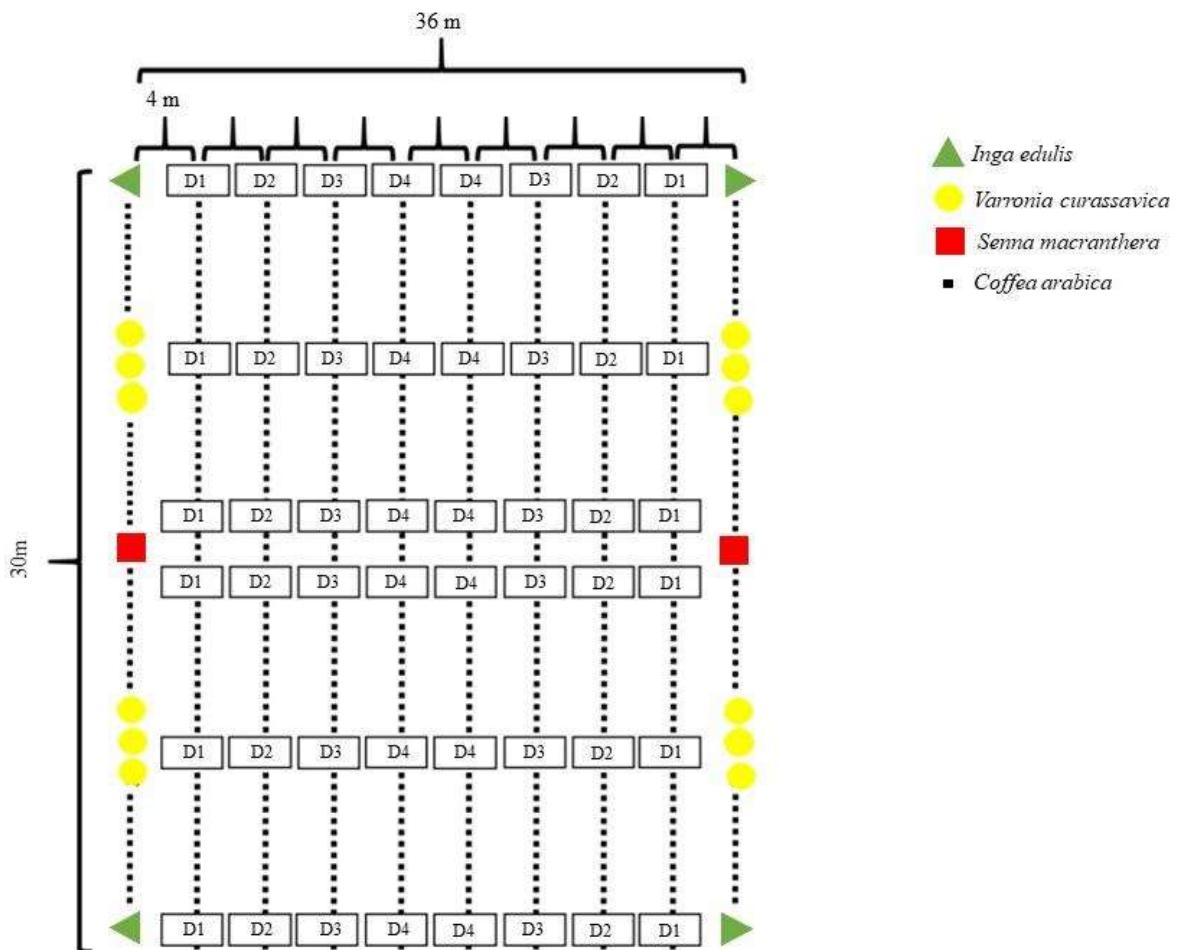


Figure 1: Adapted Botti 2021. Diversify Coffee system (1080 m²). Black points represent coffee plants, yellow circles represent *Varronia curassavica*, red squares represent *Senna macranthera* and green triangles represent *Inga edulis*. D1, D2, D3 and D4 represent the coffee plants sampled in their respective distances from IPS: D1 represents distance 1 from IPS (4 m); D2 represents distance 2 from IPS (8 m); D3 represents distance 3 from IPS (12 m) and D4 represent distance 4 from IPS (16 m).

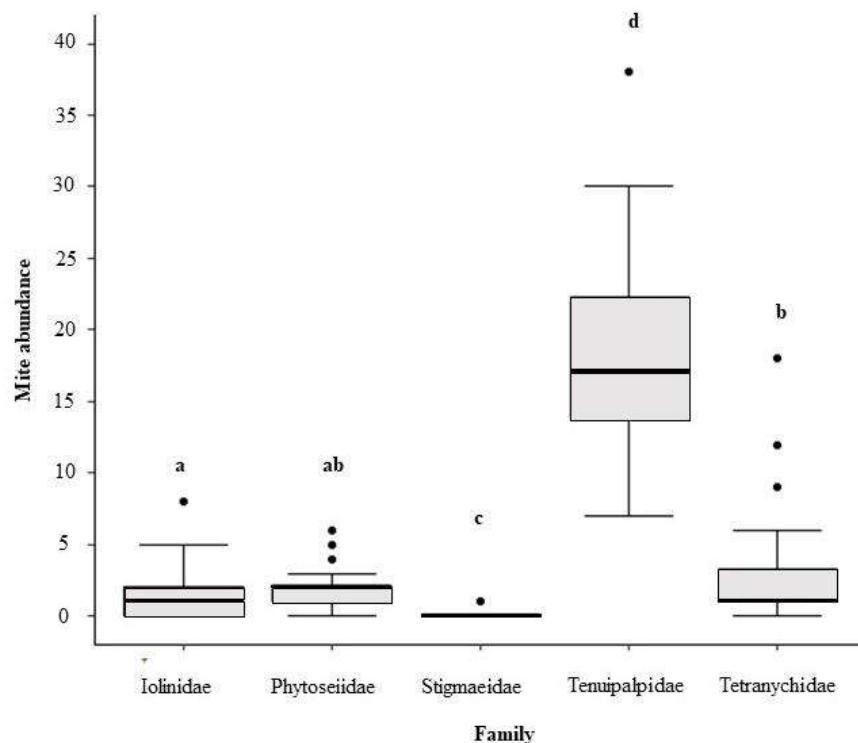


Figure 2: Abundance of mites in the coffee system. The thicker line indicates the medians. Boxes represent 25 to 75 percent of the data distribution and black spots are outliers. Different letters correspond to significant differences between treatments.

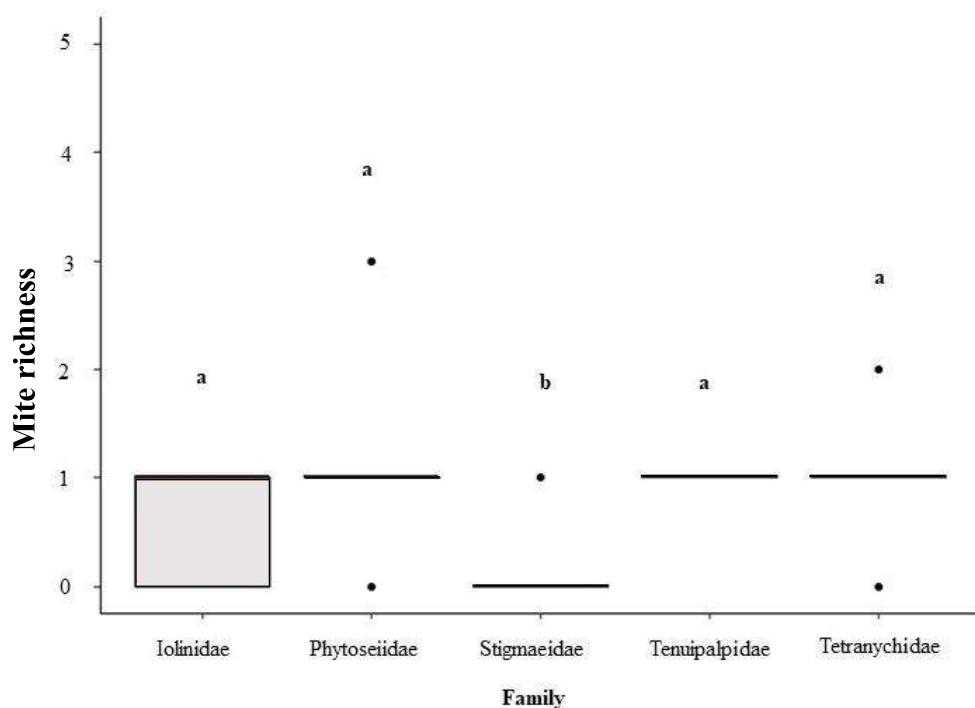


Figure 3: Richness of mites in the coffee system. The thicker line indicates the medians. Boxes represent 25 to 75 percent of the data distribution and black spots are outliers. Different letters correspond to significant differences between treatments.

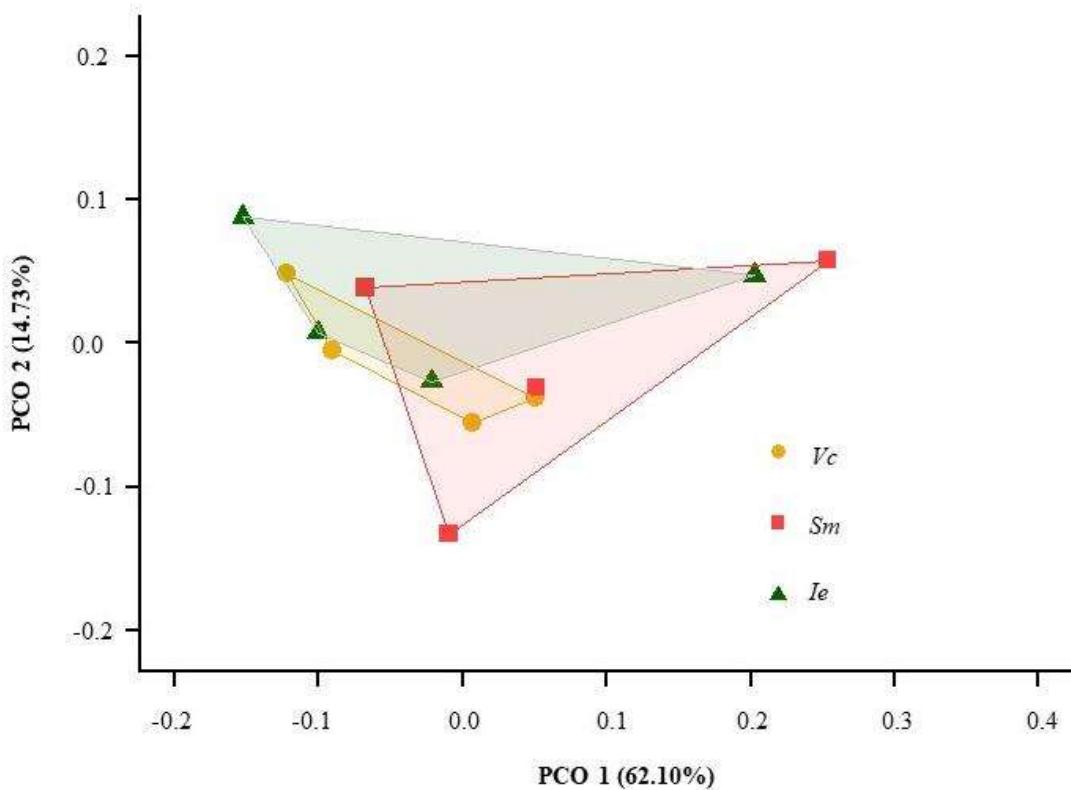


Figure 4: Principal coordinate analysis (PCO) plot based on Bray–Curtis dissimilarity index from samples of predator and phytophagous composition in coffee plants. Yellow circles represent *Varrovia curassavica* (*Vc*), red squares represent *Senna macranthera* (*Sm*) and green triangles represent *Inga edulis* (*Ie*).

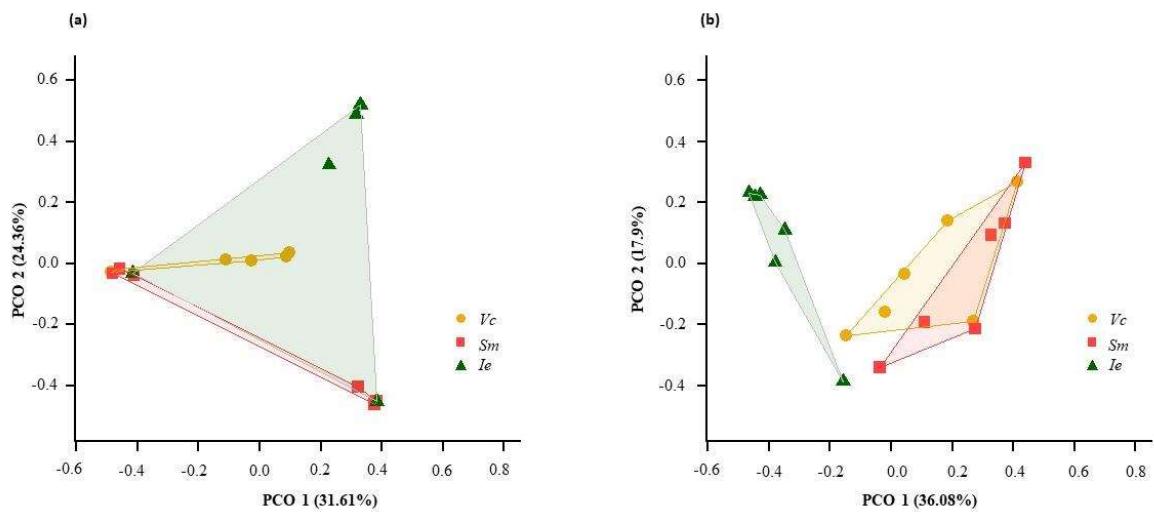


Figure 5: Principal coordinate analysis (PCO) plot based on Bray–Curtis dissimilarity index from samples of predator and phytophagous composition between IPS. Composition of phytophagous (a) and predators (b) from IPS. Yellow circles represent *Varronia curassavica* (*Vc*), red squares represent *Senna macranthera* (*Sm*) and green triangles represent *Inga edulis* (*Ie*).

Table S1: Comparison of response models to the mite abundance and richness. AICc is Akaike Information Criterion corrected; ΔAICc is the difference in AICc between the model and the model with the smallest AICc; AICcWt is the model weight according to ΔAICc .

Richness	AICc	ΔAICc	AICcWt
model 4	331,16	0,00	1,00
model 2	343,51	12,35	0,00
null model	352,46	21,30	0,00
model 3	357,19	26,03	0,00
model 1	475,45	144,29	0,00
Abundance	AICc	ΔAICc	AICcWt
model 4	698,14	0,00	0,84
model 2	701,47	3,33	0,16
model 3	711,50	13,36	0,00
model 1	798,81	100,67	0,00
null model	910,01	211,87	0,00

Chapter 2

New tydeid mite (Acari: Prostigmata) associated to coffee and inga plants

Júlia Jantsch Ferla¹; Tairis Da-Costa²; Gabriel Lima Bizarro³, Elem Fialho Martins¹, Noeli Juarez Ferla³, Madelaine Venzon⁴, Angelo Pallini¹

¹ Department of Entomology, Federal University of Viçosa, Minas Gerais, Brazil.

² Laboratório de Evolução, Ecologia Química e Quimiotaxonomia, Universidade Federal do Rio Grande do Sul, Porto Alegre, Rio Grande do Sul, Brazil.

³ Laboratório de Acarologia, Universidade do Vale do Taquari - Univates, Lajeado, Brazil.

⁴Agriculture and Livestock Research Enterprise of Minas Gerais (EPAMIG), Viçosa, MG, Brazil.

Abstract

A new species of Tydeidae (Acari: Prostigmata) is herein described and illustrated. It was collected from *Coffea arabica* L. (Rubiaceae) and *Inga edulis* Mart. (Fabaceae) plants from Cerrado biome in the state of Minas Gerais, Brazil. It has been named *Tydeus mineirensis* sp. nov.. A dichotomous key of Tydeoidea recorded on coffee crops in the states of São Paulo and Minas Gerais is provided.

Keywords: Cerrado biome, *Coffea arabica*, *Inga edulis*, mite, *Tydeus*.

Introduction

Brazil is the largest world coffee producer and exporter, being Minas Gerais the state with the largest cultivated area in the country. The main species cultivated is arabica coffee (*Coffea arabica* L.) (Conab 2021). In attempt to diversify coffee crops, leguminous trees of the genus *Inga* Miller (Fabaceae) are used by coffee farmers in some Brazilian regions (Cardoso *et al.* 2001; Grossman 2003; Souza *et al.* 2010). Besides providing shade and wood, and fixing nitrogen, these trees possess extrafloral nectaries that benefit natural enemies of coffee pests, thereby decreasing pest damage (Rezende *et al.* 2014, 2021).

Coffee crops harbor several mite species, and some of them belong to the Tydeidae family (Pallini 1991; Mineiro *et al.* 2001; Spongoski *et al.* 2005; Mineiro *et al.* 2006a, 2006b; Marchetti 2007; Mineiro *et al.* 2008, 2009, 2010; Berton *et al.* 2019; Mineiro *et al.* 2019). The superfamily Tydeoidea was reorganized by André & Fain (2000) and comprises four families: Iolinidae, Ereyntidae, Edbakerrelidae and Tydeidae. Only Edbakerrelidae was not recorded in the Brazilian coffee crops. Tydeidae is the commonest Tydeoidea family found in this crop (Pallini 1991; Mineiro *et al.* 2001; Spongoski *et al.* 2005; Mineiro *et al.* 2006a, 2006b; Marchetti 2007; Mineiro *et al.* 2008, 2009, 2010; Berton *et al.* 2019; Mineiro *et al.* 2019).

The genus *Tydeus* comprises 49 species, but only two of them are described from Brazil: *Tydeus riopardensis* Silva, Cunha & Ferla and *T. manoi* Silva, Rocha & Ferla from the state of Rio Grande do Sul (Silva *et al.* 2016). The aim of this work was to describe a new species of Tydeidae, from the state of Minas Gerais, based on specimens collected on both *C. arabica* and *I. edulis* and to provide a dichotomous key of Tydeoidea reported from coffee crops in the states of Minas Gerais and São Paulo, Brazil.

Material and Methods

Samples of *C. arabica* and *I. edulis* leaves were collected from the EPAMIG Experimental Farm, in Patrocínio county (18°59'52.0"S 46°58'59.8"W), Minas Gerais, Brazil, in the Cerrado biome, during 2020 - 2021. *Inga edulis* was intercropped with *C. arabica*, and it possesses extrafloral nectaries, which provide alternative food to the natural enemies of coffee pests and enhances conservation biological control on the

crop (Rezende *et al.* 2014; 2021). All collected leaves were placed in plastic bags, kept in a styrofoam box with Gelox® at low temperature and taken to the Laboratory of Acarology, at the Department of Entomology of Federal University of Viçosa (UFV), Viçosa, Brazil. The mites were mounted on slide glasses under a stereomicroscope in Hoyer's medium (Krantz and Walter 2009).

The identification of Tydeoidea mites was done with a phase contrast microscope and based on dichotomous keys (André 1980; Kaźmierski 1998; Silva *et al.* 2016). Drawings were made using Adobe illustrator® program. The nomenclature of idiosomal chaetotaxy follows Kaźmierski (1989, 1998) and leg chaetotaxy follows André (1981). All measurements are in micrometers (μm) and the measurements are those of the holotype followed by the range of the paratypes in parentheses. Number of setae on leg segments are from tarsi to trochanter.

The dichotomous key was made with the species of Tydeoidea sampled and those already reported on coffee plants (*C. arabica* and *C. canephora* Pierre) from the states of Minas Gerais and São Paulo (Pallini 1991; Mineiro *et al.* 2001; Spongoski *et al.* 2005; Mineiro *et al.* 2006a, 2006b; Marchetti, 2007; Mineiro *et al.* 2008, 2009, 2010; Berton *et al.* 2019; Mineiro *et al.* 2019).

Results

Systematic

Family **Tydeidae** Kramer 1877

Subfamily **Tydeinae** Kramer sensu André 1980

Genus ***Tydeus*** (Koch 1835)

Tydeus mineirensis Ferla & Pallini sp. nov.

(Figures 1 - 5)

General diagnosis

All dorsal setae, except for *ro*, *la* and *bo*, are short and spatulated. Ventral setae are slender and smooth. Dorsal ornamentation is striated, type *Tydeus*. Some specimens with and others without longitudinal striae between *ro*, *la* and *bo*. Setae *d* are bifurcated. Some seta on legs I-IV are smooth and some serrated. Empodium with two hooks.

Description. Adult female (n=7)

Dorsum (Figure 1) - Dorsum with 13 pairs of setae (*ro*, *la*, *bo*, *ex*, *c1*, *c2*, *d*, *e*, *f1*, *f2*, *h1*, *h2* and *ps1*). Dorsal shield (excluding gnathosoma) 280 (268 - 300), width 203 (200 - 210). Setae *ps1* is located dorsally. Dorsum of idiosoma faintly striated. Dorsal setae are short and spatulate, except for *ro* and *la*, that are slightly serrated and *bo* is smooth. Striation U-shaped between setae *c1* and *d1*, transverse to diagonal posterior to setae *d1*. There is a variation in this species, some specimens have a suture between the propodosoma and the hysterosoma and some of them do not.

Lengths of dorsal setae: *ro* 15 (13 - 15), *la* 15 (13 - 15), *bo* 33 (33 - 35), *ex* 15 (10 - 15), *c1* 13 (10 - 13), *c2* 10 (10 - 13), *d* 15 (10 - 15), *e* 13 (10 - 15), *f1* 15 (13 - 15), *f2* 13 (10 - 15), *h1* 13 (13 - 15) and *h2* 15. Distances between dorsal setae: *ro-ro* 30 (23 - 33), *la-la* 88 (88 - 128), *bo-bo* 60 (58 - 68), *ex-ex* 138 (53 - 70) *c1-c1* 48 (48 - 55), *c2-c2* 175 (170 - 210), *d1-d1* 33 (33 - 38), *e1-e1* 90 (88 - 100), *f1-f1* 23 (23 - 28), *f2-f2* 58 (55 - 65), *h1-h1* 23 (20 - 25), *h2-h2* 50 (38 - 50), *ro-la* 33 (30 - 40), *c1-d1* 50 (48 - 53), *d1-e1* 40 (23 - 40), *e1-f1* 43 (38 - 43), *f1-f2* 15 (13 - 18) and *h1-h2* 13(13 - 15).

Venter (Figure 2) - All venter striated (Coxa I, III and IV with reticulations). All ventral setae are slender and smooth. Anogenital area with five pairs of genital setae, four pairs of aggenital and one pair of pseudoanal setae. Genital striation pattern extended. Shape of *cg* organ elliptical. Epimeral formula: 2-1-3-1.

Measurements of setae: *pt* 13 (10 - 15), *mt α* 13(10 - 15) and *mt β* 13 (10 - 13). Four pairs of aggenital setae (*ag1*, *ag2*, *ag3* and *ag4*) and one pair of pseudanal setae (*ps2*). Setal lengths: *ag1* 10 (5 - 10), *ag2* 10 (5 - 10), *ag3* 13 (5 - 13), *ag4* 8 (8 - 13), *g1* 5 (5 - 8), *g2* 5 (4 - 8), *g3* 6 (5 - 6), *g4* 5 (5 - 6), *g5* 7 (5 - 10) and *ps2* 7 (7 - 13).

Gnathosoma (Figure 3). Length 48 (43 - 53), width 57 (50 - 58). Gnathosoma visible from above. Setal lengths: *sc1* 14 (8 - 14), *sc2* 9 (9 - 18). Palp (from genus to tarsus) 44 (38 - 60) long, setation 5(+1 ω)-2-2 (Figure 4), setae *d* distally bifurcated. Palp tarsus with eupathidium $\rho\zeta$ 44 (43 - 45) semilunar distally. Palptarsus significantly longer than cheliceral digits. Cheliceral stilettos 14 (13 - 15) long.

Legs. (Figure 5A-D) - Leg setae are smooth, except for some seta that are serrated: two dorsal setae from genu I, two ventral from genu II and one dorsal from genu III and IV each, two on tibia IV and one dorsal on femur I. Tarsi I-IV with two claws and hairy empodium but empodial hooks (*om*) are absent. Some leg segments with ventral reticulations. Chaetotaxy of legs I-IV (tarsus to trochanter): I: 8(+1 ω)-3(+1 κ)-3-3-1 (Figure 5A); II: 6(+1 ω)-2-2-2-0 (Figure 5B); III: 5-2-1-1-1 (Figure 5C) and; IV: 5-2-1-1-0 (Figure 5D). Length of leg (from trochanter to tarsi): I 133 (123 - 155), leg II 113 (88 - 113), leg III 130 (103 - 130) and leg IV 143 (123 - 143). Length of tarsus + apotele I 44 (43 - 45) and 10 (10 - 13) width, length of solenidion ω 5 (4 - 5), length of seta f' 20 (11 - 20), f'' 13 (11 - 21), length of seta k 3 (1-3), length of ωll 3 (2 - 3).

Male. Unknown.

Differential diagnosis

The new species is similar to *T. munsteri* Meyer & Ryke, *T. nieuwkerkeni* André, *T. taiwanensis* (Baker) and *T. costensis* Baker. However, it differs from the number of genital setae: *T. munsteri* presents 4 setae, the others species 6 and *Tydeus mineirensis sp. nov.* 5 setae.

The dorsal setae of *T. munsteri* is similar to *Tydeus mineirensis sp. nov.*, while the dorsal setae of *T. nieuwkerkeni* is characterized by three pairs of short and spatulated setae. *Tydeus taiwanensis* has some dorsal setae slightly spatulated and others broadly and *T. costensis* has dorsal body setae slender and slightly serrated. Besides these characteristics, *T. taiwanensis*, *T. costensis* and *T. munsteri* have *bo* not exceeding the base of the next arrow (*c1*), while *bo* of the new species exceed the base of *c1*.

Etymology

The species name is a tribute for the first Tydeidae described for state of Minas Gerais.

Type material

Female holotype and 6 female paratypes.

Patrocínio, Minas Gerais, Brazil, ($18^{\circ}59'52.0"S$ $46^{\circ}58'59.8"W$), collected by Ferla J. J. from Jun. 2020 to Feb. 2021. Holotype and three paratype are deposited in the collection of the Acarology and Entomology Department, Escola Superior de Agricultura “Luiz de Queiroz”, University of São Paulo (ESALQ/USP), Piracicaba, São Paulo State, Brazil. There are one paratype deposited at Laboratory of Acarology, Department of Entomology, Federal University of Viçosa (UFV), Viçosa, Minas Gerais state, Brazil and two paratypes deposited at the Museu de Ciências Naturais (ZAUMCN), Universidade do Vale do Taquari - Univates, Lajeado, Rio Grande do Sul state, Brazil.

Dichotomous key of Tydeoidea mites on Coffee plants (*C. arabica* and *C. canephora*) to São Paulo and Minas Gerais state, Brazil (based partially on André 1980; André and Fain 2000; Baker 1965; Kaźmierski 1998; Silva et al. 2016)

1. With posterior bothridia on the opisthosoma.....Ereynetidae
 - Without posterior bothridia on the opisthosoma.....2
-
2. Pretarsus of leg I present and normal
 -Tydeidae.....3
 - Pretarsus of leg I with vestigial claws or with no claws.....Iolinidae.....6
-
3. Genu II with one or two seta; double or triple eupathidia at the end of the palptarsus.....Tydeinae.....4
 - No setae on genu II; triple eupathidia at the end of the palptarsus.....Pretydeinae.....Trochanter II with seta.....*Pretydeus* André
-
4. Femur III with two setae.....*Lorryia* sensu André.....Dorsal idiosomal setae nude; reticulate areas are divided into Ac2, A(c1), Ac2, Ae1, A(d1), and Ae1
 -*Lorryia formosa* Cooreman
 - Femur III with one seta 5

5. Femur II with three setae.....*Pseudolorryia* Kaźmierski
 - Femur II with two setae.....*Tydeus* Koch..... Genital with 5 setae (g1-g5); dorsal setae spatulated, except for *ro*, *la* and *bo*
*Tydeus mineirensis* sp. nov.
6. Setae *la* reduced or absent.....*Parapronematus* Baker.....Femur III and IV with forked setae*Parapronematus acaciae* Baker
 -Setae *la* present and normal.....7
7. Tarsus I with 8 setae; femur IV divided.....*Pseudopronematulus* Fan & Li.....Tarsus IV with five setae and tarsus II with six setae.....*Pseudopronematulus nadirae* Silva, Da-Costa & Ferla
 -Tarsos I with 8 setae; femur IV entire.....8
8. Tarsus III and IV with 6 setae; four pairs of aggenital setae*Homeopronematus* André
 -Tarsus III with 6 setae and tarsus IV with 5 setae; three pairs of aggenital setae*Metapronematus* André

Acknowledgements

The authors are grateful to André Costa Cardoso, Milena Kalile, Pedro Hermano Marques Gonçalves Nascimento and Shauanne Dias Pancieri for helping in the slides mounting and to Wesley Borges Wurlitzer for taxonomic help and suggestions. This study was financed in part by Coordination for the Improvement of Higher Education Personnel (CAPES) finance code 001, Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Consórcio Brasileiro de Pesquisa e Desenvolvimento do Café (CBP & D-Café) and Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG).

References

- André H.M. 1980. A generic revision of the family Tydeidae (Acari: Actinedida). IV. Generic descriptions, keys and conclusion. Bull. ann. Soc. r. belge entomolog., 116: 103–168.
- André H.M. 1981. A generic revision of the family Tydeidae (Acari: Prostigmata). III. Organotaxy of the legs. Acarologia, 22, 165–178.
- André H.M., Fain A. 2000. Phylogeny, ontogeny and adaptive radiation in the superfamily Tydeoidea (Acari: Actinedida), with a reappraisal of morphological characters. Zool J Linnean Soc, 130: 405-448. 10.1006/zjls.1999.0237
- Baker E.W. 1965. A review of the genera of the family Tydeidae (Acarina). Advances in Acarology, 2: 95–133.
- Berton L.H.C., Mineiro J.L.C., Sato M.E., Joaquim A.D.A.F., Raga A. 2019. Mite fauna of a coffee agroecosystem (*Coffea arabica* L.) in the municipality of Monte Alegre do Sul, São Paulo State, Brazil. Part I. Acarologia, 59(4): 542-550. 10.24349/acarologia/20194352
- Cardoso I.M., Guijt I., Franco F.S., Carvalho A.F., Neto P.F. 2001. Continual learning for agroforestry system design: university, NGO and farmer partnership in Minas Gerais, Brazil. Agric. Syst., 69(3): 235-257. [https://doi.org/10.1016/S0308-521X\(01\)00028-2](https://doi.org/10.1016/S0308-521X(01)00028-2)
- Conab. 2021. Companhia Nacional de Abastecimento. Acompanhamento da safra brasileira de café, Safra 2021, Segundo levantamento, Brasília, 8(2): 1-63.
- Grossman J.M. 2003. *Inga oerstediana* in organic coffee agroecosystems: Nitrogen fixation, nodulation and farmer knowledge of soil processes in Chiapas, Mexico [PhD Thesis]. University of Minnesota: Faculty of the graduate school of the University of Minnesota.

Kazmierski A. 1998 Tydeinae of the world. Generic relationships, new & redescribed taxa and keys to all species. A revision of the subfamilies Pretydeinae & Tydeinae (Acari: Actinedida, Tydeidae) Part IV. *Acta Zool. Cracov.*, 41(2): 283-455.

Kaźmierski A. 1989. Morphological studies on Tydeidae (Actinedida; Acari). I. Remarks about the segmentation, chaetotaxy and poridotaxy of idiosoma. *Acta Zool. Cracov.*, 32: 69–83.

Koch C.L. 1835. Deutsche Crustacea, Myriapoda, Arachnida, 4.

Kramer P. 1877. Tydidae. *Archiv für Naturgeschichte*, 43 p. 232–246.

Krantz G.W., Walter D.E. 2009. A manual of acarology. Third Edition. USA, Texas Tech University Press. pp. 807.

Marchetti M.M. 2007. Ácaros do cafeiro em Minas Gerais com chave de identificação [Master dissertation]. Viçosa: Universidade Federal de Viçosa. pp 159.

Mineiro J.L.D.C., Raga A., Sato M.E., Matioli A.L., Berton L.H.C. 2010. Ácaros de cafeiro (*Coffea* spp.) no Estado de São Paulo, Brasil: Parte II. Prostigmata. *Biota Neotrop.*, 10(4): 215-225. <https://doi.org/10.1590/S1676-06032010000400027>

Mineiro J.L.D.C., Sato M.E., Matioli A.L., Raga A. 2009. Ácaros plantícolas e edáficos em agroecossistema cafeiro, em Atibaia, estado de São Paulo. In: VI Simpósio de Pesquisa dos Cafés do Brasil.

Mineiro J.L.C., Sato M.E., Raga A. 2019. Domácia e ácaros em cafeeiros. *Biológico*, 81: 1-15. [10.31368/1980-6221v81a10011](https://doi.org/10.31368/1980-6221v81a10011)

Mineiro J.D.C., Sato M.E., Raga A., Arthur V., Cangani K.G., Barbosa F.V. 2006a. Diversidade de ácaros (Arachnida: Acari) em cinco cultivares de duas espécies de cafeeiros (*Coffea* spp.) em Garça, Estado de São Paulo. *Arq. Inst. Biol.*, 73(3): 333-341.

Mineiro J.L.D.C., Sato M.E., Raga A., Arthur V., de Moraes G.J., Sarreta F.D.O., Carrijo A. 2006b. Diversidade de ácaros (Arachnida: Acari) em *Coffea arabica* L. cv. Mundo Novo, nos municípios de Jeriquara e Garça, Estado de São Paulo. *Biota Neotrop.*, 6: 333-341. <https://doi.org/10.1590/S1676-06032006000200012>

Mineiro J.D.C., Sato M.E., Raga A., Souza M.F.D., Siloto R.C., de Moraes G.J., Spongoski S. 2001. Distribuição da acarofauna em cafeiro (*Coffea arabica* var. Catuaí amarelo), em Atibaia, SP. II Simpósio de Pesquisa dos Cafés do Brasil. p. 1921-1926.

Mineiro J.L.D.C., Sato M.E., Raga A., Souza M.F.D., Spongoski S. 2008. Incidência de ácaros em cafeiro cv. Catuaí Amarelo. *Bragantia*, 67(1): 197-201.

Pallini A. 1991. Acarofauna e predação de ácaros fitófagos por ácaros predadores em cafeiro (*Coffea arabica* L.) no Sul de Minas Gerais [Master dissertation]. Lavras: Universidade Federal de Lavras. pp 91.

Rezende M.Q., Venzon M., Perez A.L., Cardoso I.M., Janssen A. 2014. Extrafloral nectaries of associated trees can enhance natural pest control. *Agric. Ecosyst. Environ.*, 188: 198-203. <https://doi.org/10.1016/j.agee.2014.02.024>

Rezende, M.Q., Venzon, M., dos Santos, P.S., Cardoso, I.M. & Janssen, A. (2021). Extrafloral nectary-bearing leguminous trees enhance pest control and increase fruit weight in associated coffee plants. *Agric. Ecosyst. Environ.*, 319: 107538. <https://doi.org/10.1016/j.agee.2021.107538>

Silva G.L., Metzelthin M.H., Da Silva O.S., Ferla N.J. 2016. Catalogue of the mite family Tydeidae (Acari: Prostigmata) with the world key to the species. *Zootaxa*, 4135 (1): 1-68. <http://doi.org/10.11646/zootaxa.4135.1.1>

Souza H.N., Cardoso I.M., Fernandes J.M., Garcia F.C., Bonfim V.R., Santos A.C., Carvalho A.F., Mendonça E.S. 2010. Selection of native trees for intercropping with coffee in the Atlantic Rainforest biome. *Agrofor. Syst.*, 80: 1-16. <https://doi.org/10.1007/s10457-010-9340-9>.

Spongoski S., Reis P.R., Zacarias M.S. 2005. Acarofauna da cafeicultura de cerrado em Patrocínio, Minas Gerais. *Ciênc. agrotec.*, 29: 9-17. <https://doi.org/10.1590/S1413-7054200500010000>

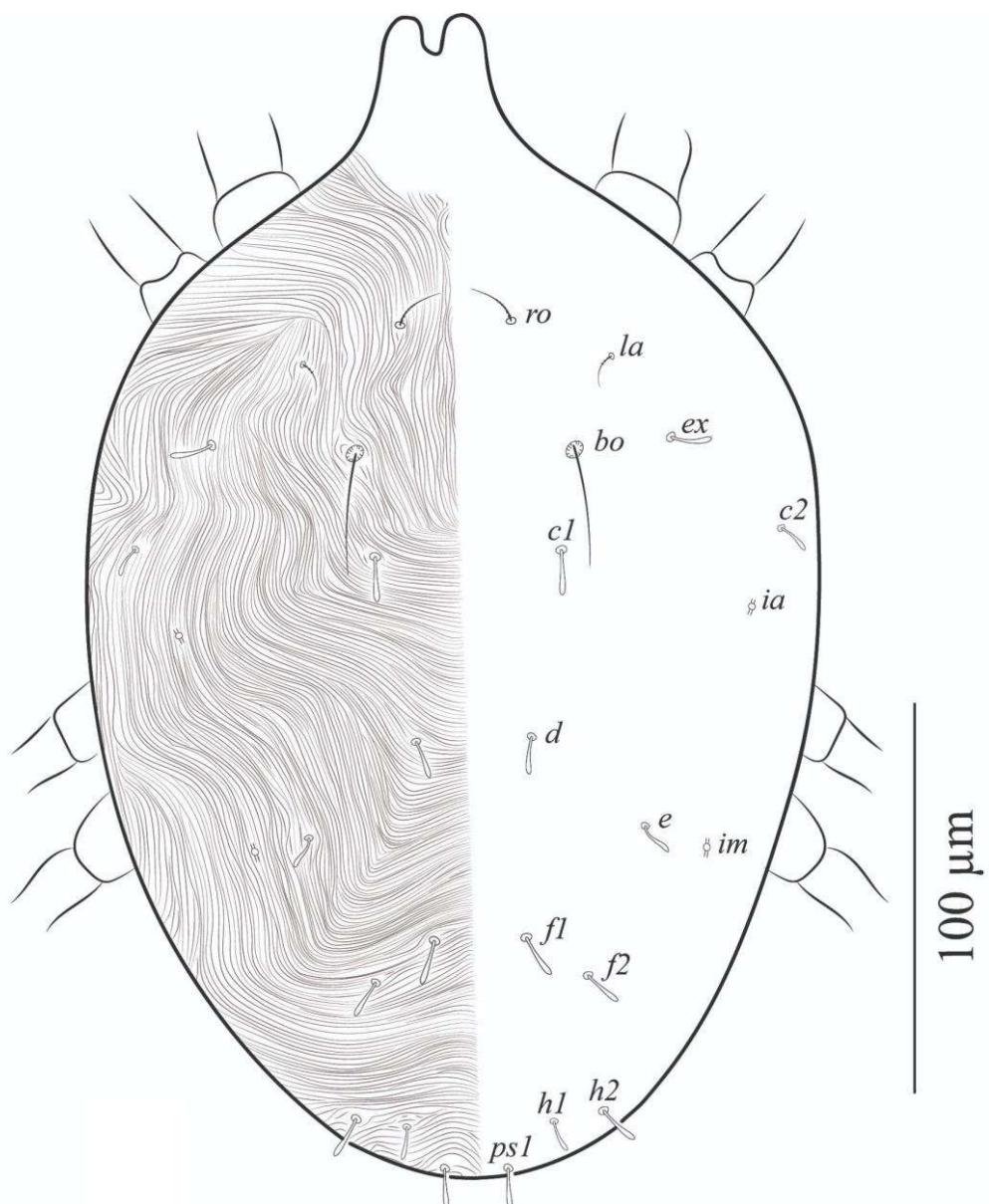


Figure 1: *Tydeus mineirensis* sp. nov. Female. Idiosoma in dorsal view.

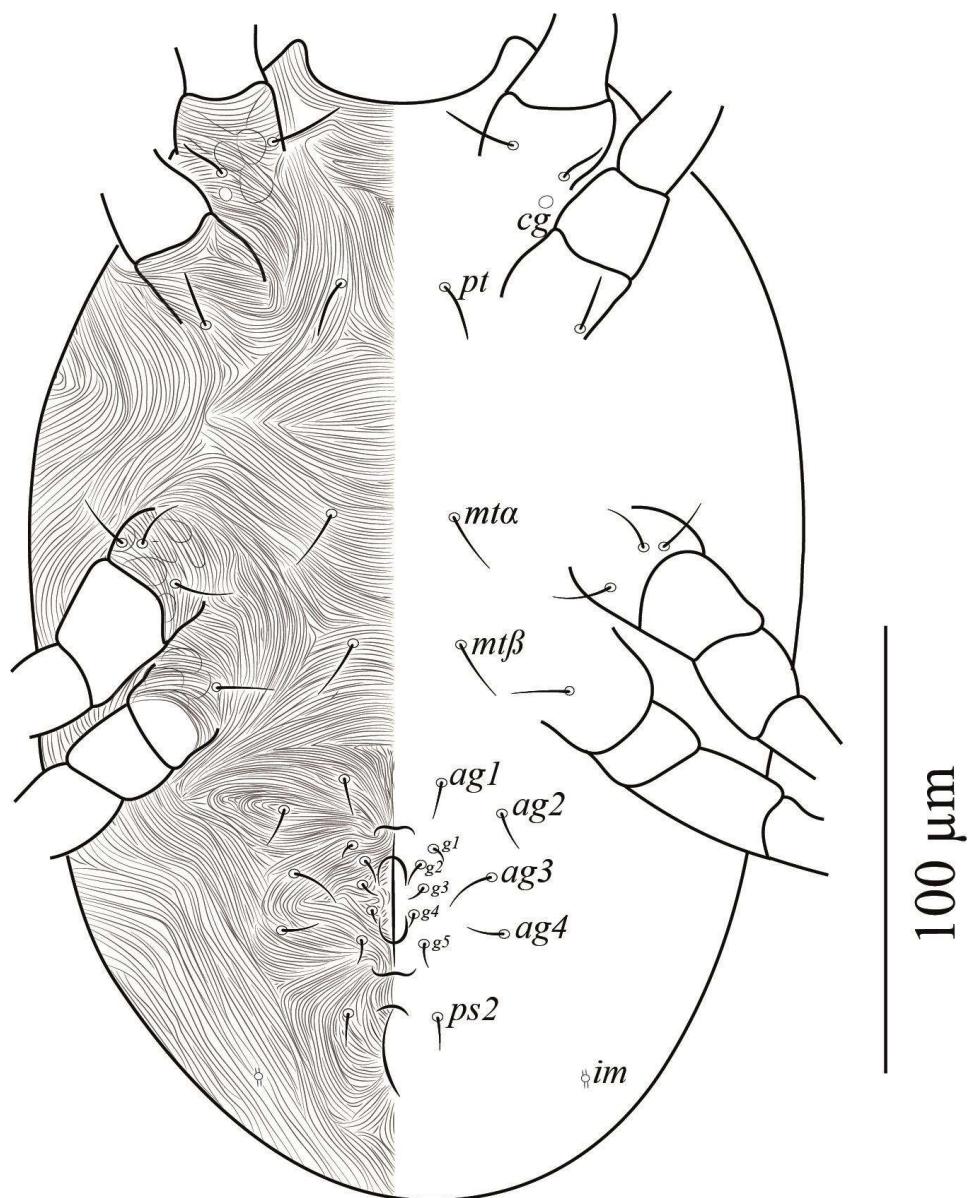


Figure 2: *Tydeus mineirensis* sp. nov. Female. In ventral view.

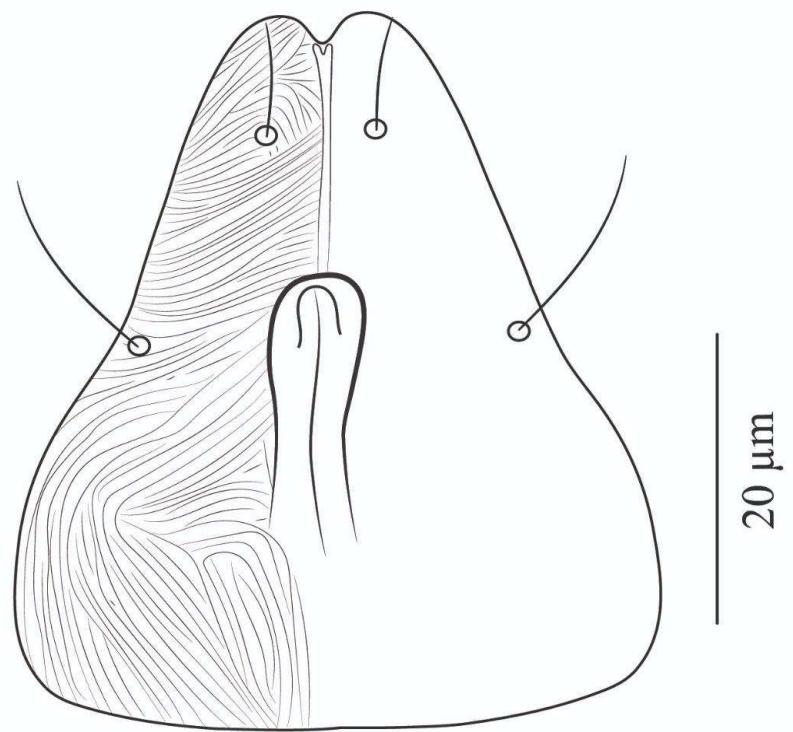


Figure 3: *Tydeus mineirensis* sp. nov. Female. Gnathosoma in ventral view.

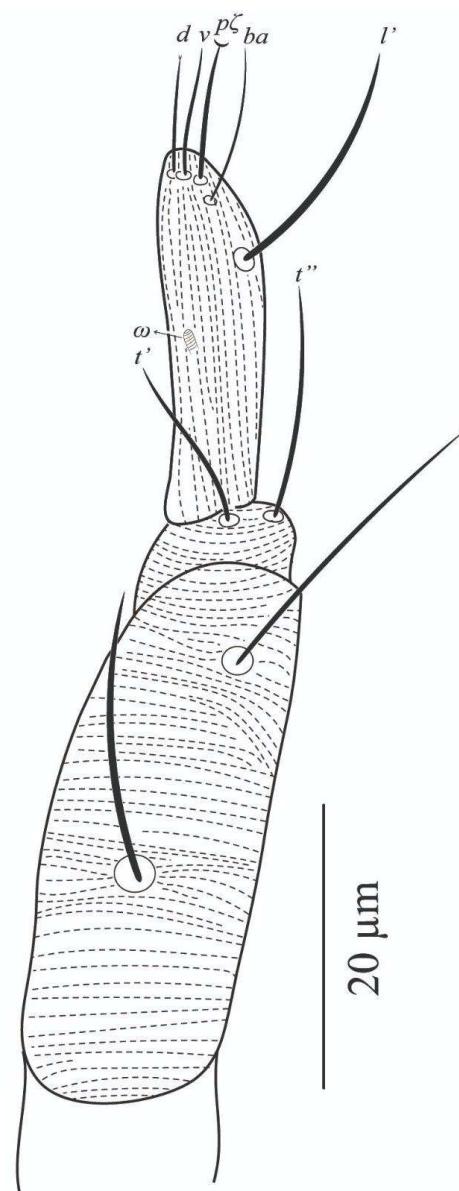


Figure 4: *Tydeus mineirensis* sp. nov. Female. Palptarsus.

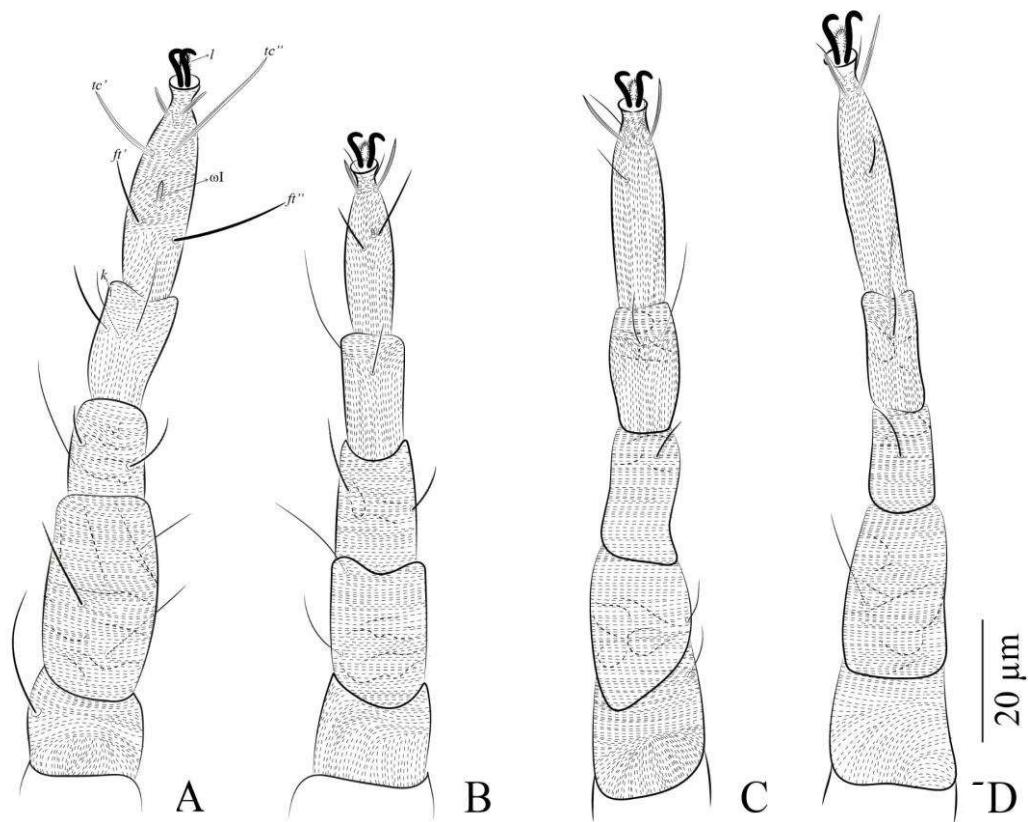


Figure 5: *Tydeus mineirensis* sp. nov. Female. A - Leg I; B - Leg II; C - Leg III; D - Leg IV; (From trochanter to tarsi).

GENERAL CONCLUSIONS

Our main conclusions provide evidence that the abundance and richness of predatory and phytophagous mites families are different in coffee intercropped with plants that provides alternative food, but their composition is not. Furthermore, the communities of these mites do not change if their distance from the IPS increases.

The composition of phytophagous and predatory mites is different in IPS. It is similar between *S. macranthera* and *V. curassavica* and different from *I. edulis*. These 3 plant species are a reservoir of natural enemies of predatory mites and harbor herbivorous mites that are not pests on coffee and they can serve as alternative prey to predators in the crop.

Finally, we described a new mite species named *Tydeus mineirensis* sp. nov. recorded on coffee and *I. edulis* plants and it is the first Tydeidae described for Minas Gerais state.